

# **Chapter 3**

## **Bacteria and Archaea**

# **Bacterial and Archaea Structure and Function**

- **prokaryotes differ from eukaryotes in size and simplicity**
  - most lack internal membrane systems
  - term prokaryotes is becoming blurred
  - this text will use **Bacteria and Archaea**
- **prokaryotes are divided into two taxa**
  - *Bacteria* and *Archaea*

# Size, Shape, and Arrangement

- **shape**
  - **cocci and rods most common**
  - **various others**
- **arrangement**
  - **determined by plane of division**
  - **determined by separation or not**
- **size - varies**

# Shape and Arrangement-1

- **cocci (s., coccus) – spheres**
  - **diplococci (s., diplococcus) – pairs**
  - **streptococci – chains**
  - **staphylococci – grape-like clusters**
  - **tetrads – 4 cocci in a square**
  - **sarcinae – cubic configuration of 8 cocci**

# Shape and Arrangement-2

- **bacilli (s., bacillus) – rods**
  - **coccobacilli – very short rods**
- **vibrios – resemble rods, comma shaped**
- **spirilla (s., spirillum) – rigid helices**
- **spirochetes – flexible helices**

# Figure 3.1

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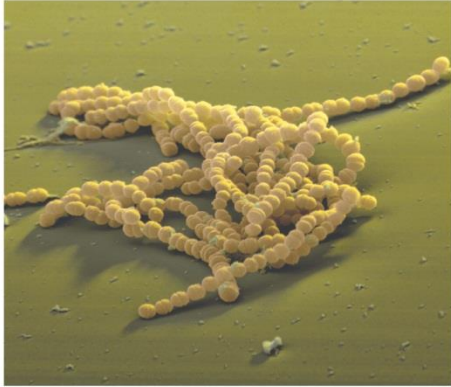
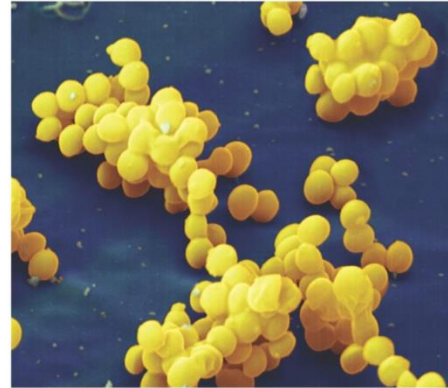


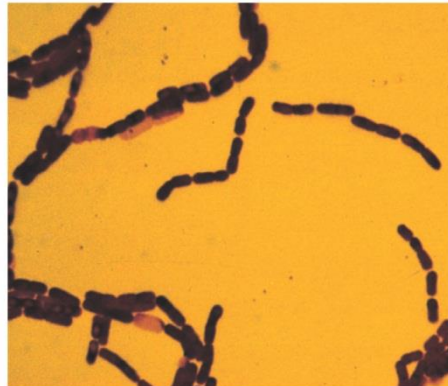
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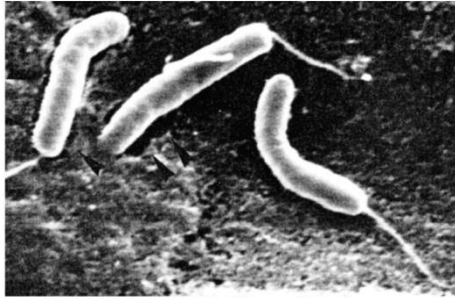
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# Shape and Arrangement-3

- **mycelium – network of long, multinucleate filaments**
- **pleomorphic – organisms that are variable in shape**
- ***Archaea***
  - **pleomorphic, branched, flat, square, other unique shapes**

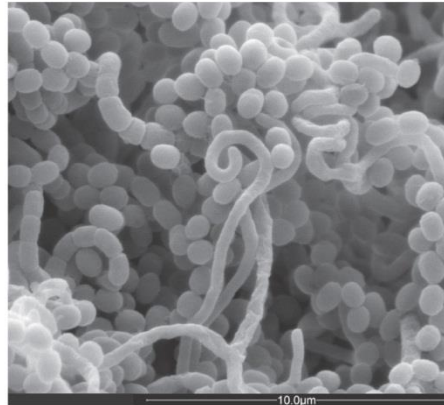
# Figure 3.2; e.g v.cholera, Streptomyces, leptospirae etc

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Centers for Disease Control

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(d) *Streptomyces*—a filamentous bacterium  
Dr. Amy Gehring

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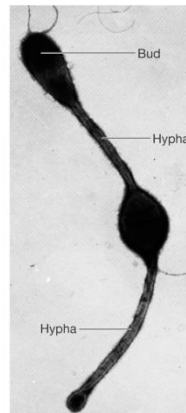
From Walther Stoeckenius: Walsby's Square Bacterium Fine Structures of an Orthogonal Prokaryote

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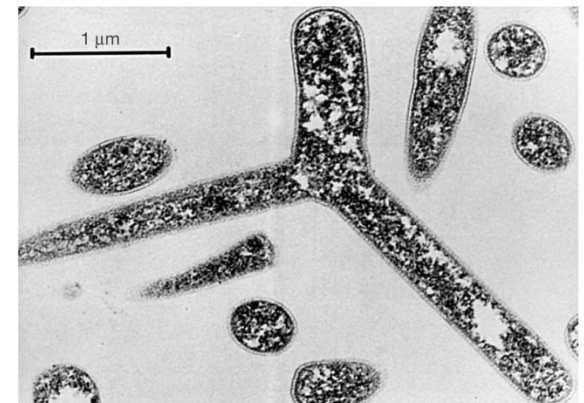
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(e) *Hyphomicrobium*

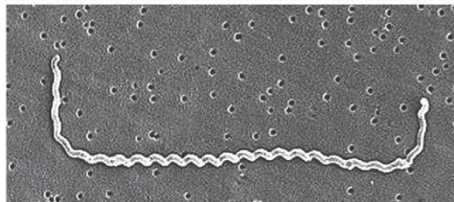
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(f) *Thermoproteus tenax*—a branched archaeal cell  
From J.T. Staley, M.P. Bryant, N. Plenning and J.G. Holt (Eds.), Bergey's Manual of System-atic Bacteriology, Vol. 3. © 1989 Williams and Wilkins Co., Baltimore, Robinson, Dept. of Micro, U. of Cal., LA

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(c) *Leptospira interrogans*—a spirochete  
CDC/NCID/HIP/Janice Carr

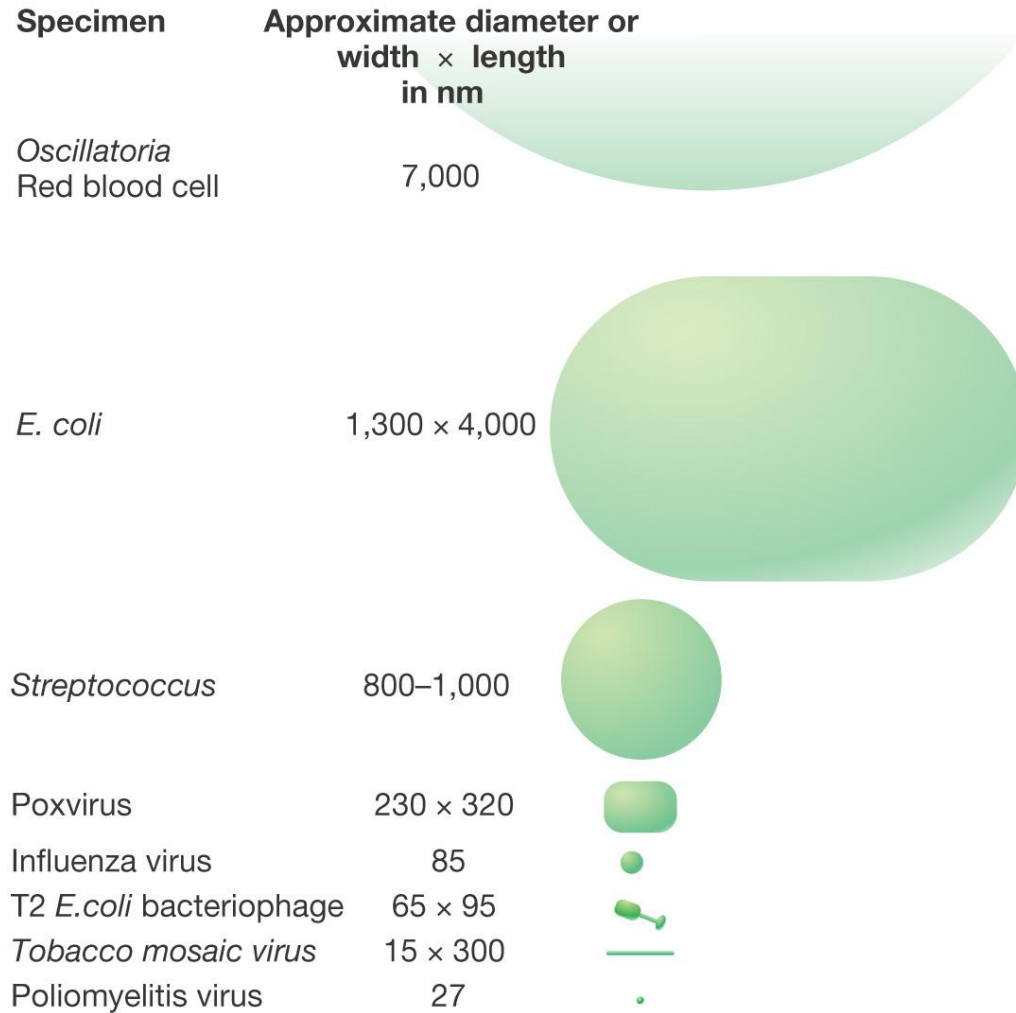


# Size

- **smallest – 0.3 (*Mycoplasma*)**
- **average rod – 1.1 - 1.5 x 2 – 6  $\mu\text{m}$   
(*E. coli*)**
- **very large – 600 x 80  $\mu\text{m}$   
*Epulopiscium fishelsoni***

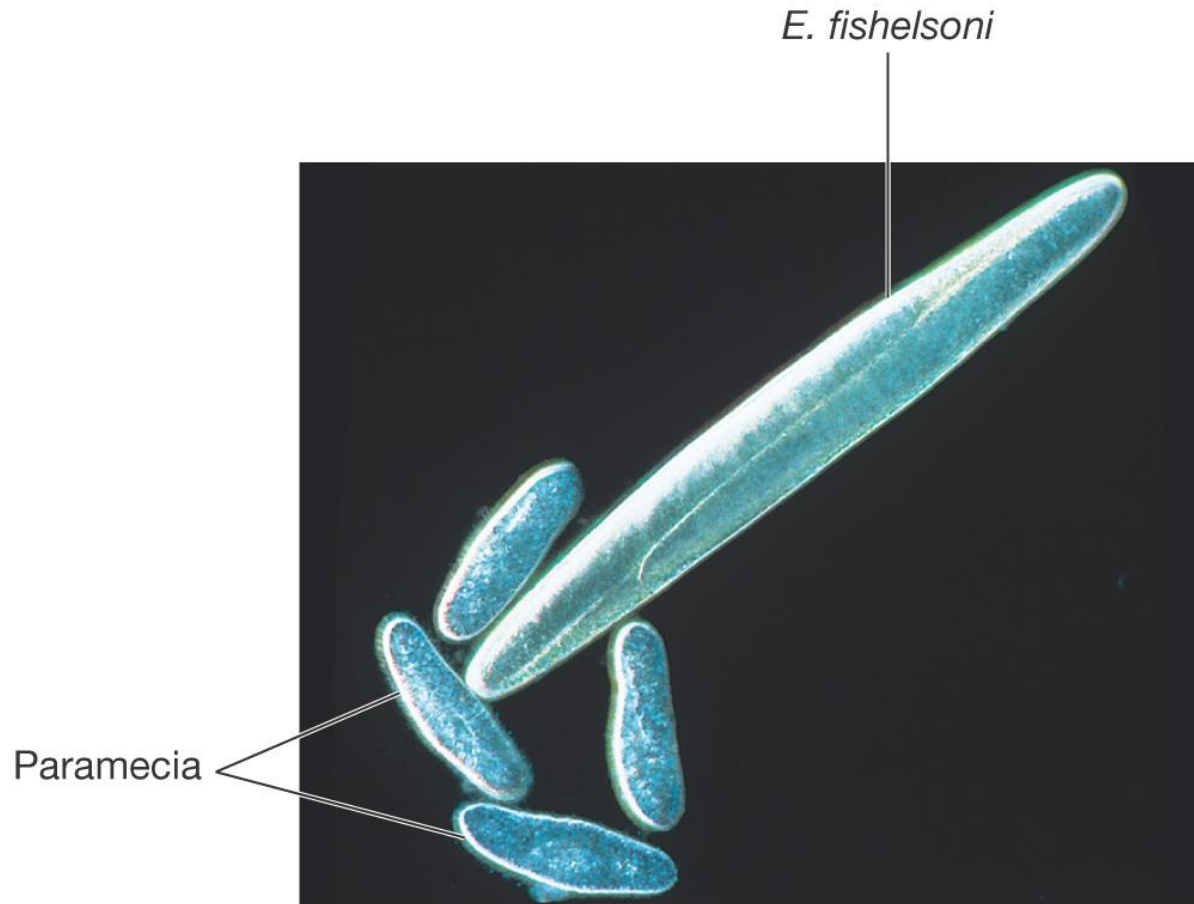
# Figure 3.3

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# Figure 3.4; *Epulopiscium fishelsoni*

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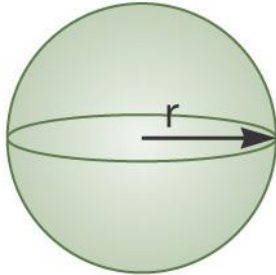
Dr. Leon J. LeBeau

# **Size – Shape Relationship**

- **important for nutrient uptake**
- **Small have higher surface to volume ratio (S/V)**
- **small size may be protective mechanism from predation**

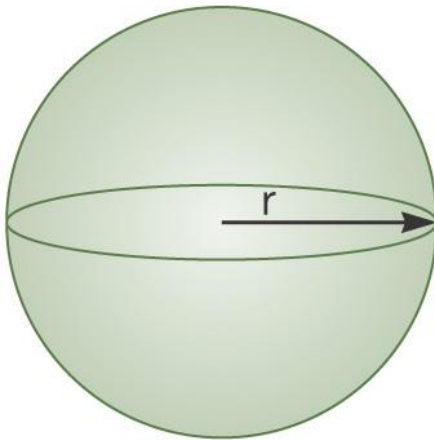
# Figure 3.5

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$$r = 1 \mu\text{m}$$
$$\text{Surface area} = 12.6 \mu\text{m}^2$$
$$\text{Volume} = 4.2 \mu\text{m}^3$$

$$\frac{\text{Surface}}{\text{Volume}} = 3$$



$$r = 2 \mu\text{m}$$
$$\text{Surface area} = 50.3 \mu\text{m}^2$$
$$\text{Volume} = 33.5 \mu\text{m}^3$$

$$\frac{\text{Surface}}{\text{Volume}} = 1.5$$

# **Cell Organization**

## **Archaea and Bacteria**

### **Common Features**

**Cell envelope – 3 layers**

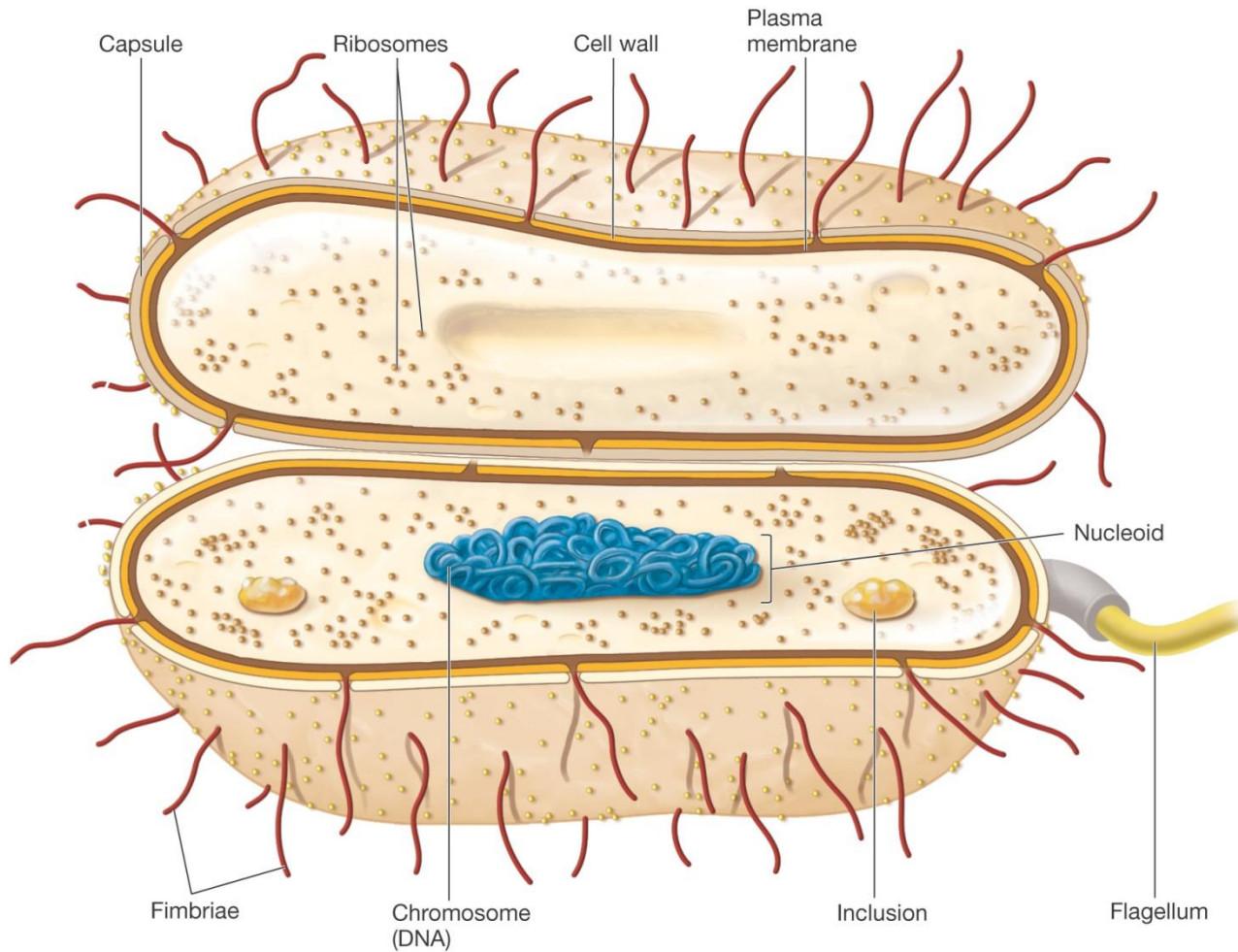
**Cytoplasm**

**External structures**

# Table 3.1

# Figure 3.6

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# **Bacterial Cell Envelope**

**Plasma membrane**

**Cell wall**

**Layers outside the cell wall**

# Bacterial Plasma Membrane

- **absolute requirement for all living organisms**
- **some bacteria also have internal membrane systems**

# Plasma Membrane Functions

- **encompasses the cytoplasm**
- **selectively permeable barrier**
- **interacts with external environment**
  - **receptors for detection of and response to chemicals in surroundings**
  - **transport systems**
  - **metabolic processes**

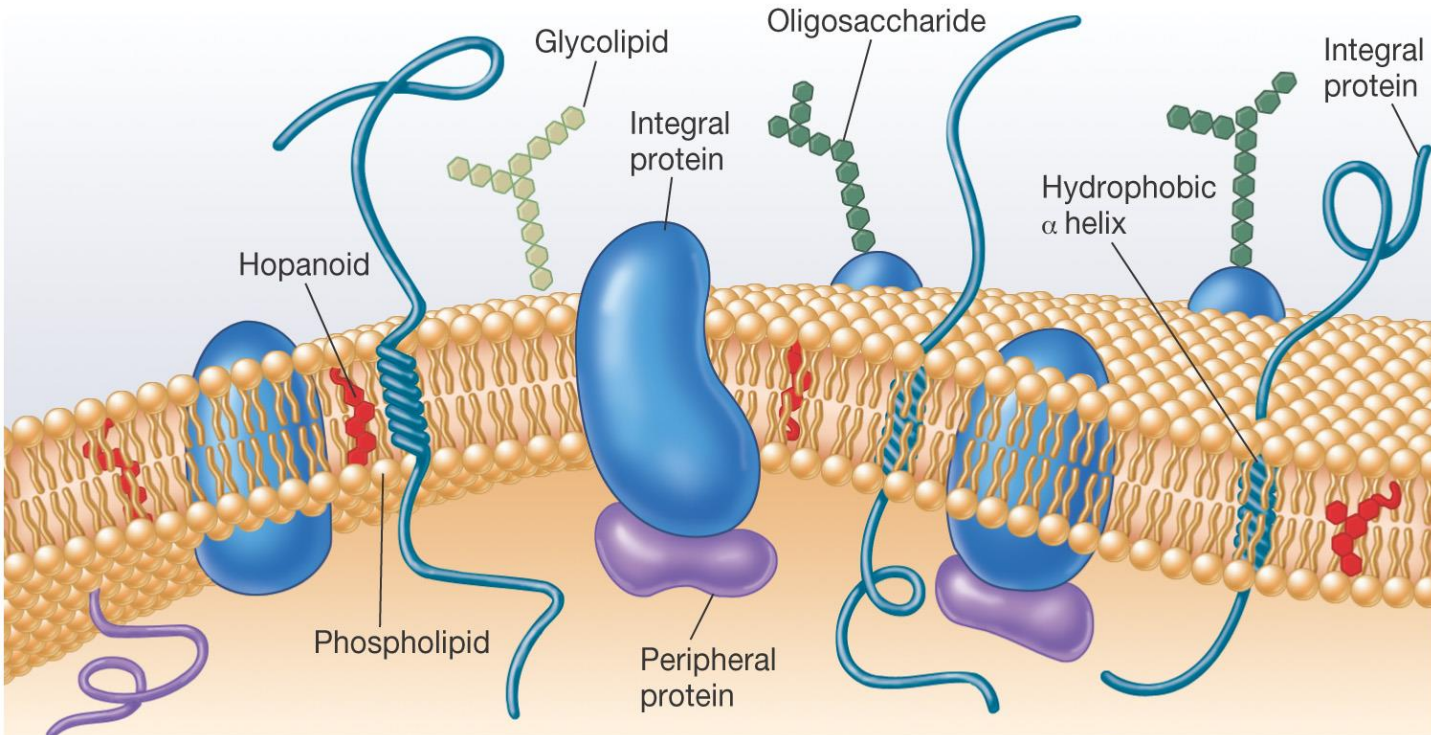
# **Fluid Mosaic Model of Membrane Structure**

- **lipid bilayers with floating proteins**
  - **amphipathic lipids**
    - **polar ends (hydrophilic – interact with water)**
    - **non-polar tails (hydrophobic – insoluble in water)**
  - **membrane proteins**

# Fluid Mosaic Model of Membrane Structure

## Figure 3.7

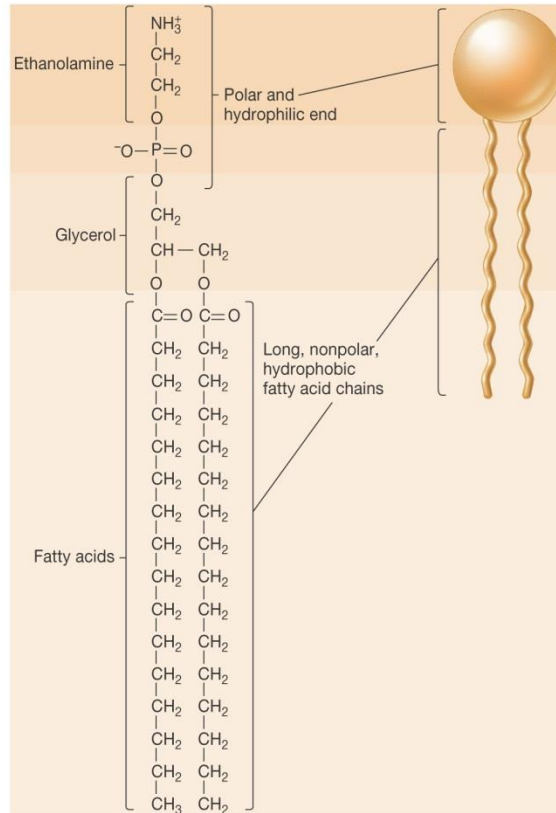
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# The Asymmetry of Most Membrane Lipids

## Figure 3.8

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# Membrane Proteins

- **peripheral**
  - loosely connected to membrane
  - easily removed
- **integral**
  - amphipathic – embedded within membrane
  - carry out important functions
  - may exist as microdomains

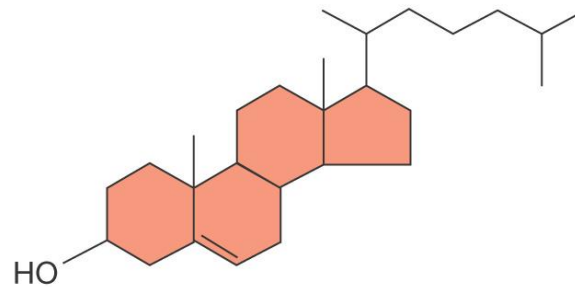
# Bacterial Lipids

- **saturation levels of membrane lipids reflect the environmental conditions such as temperature**
- **bacterial membranes lack sterols but do contain sterol-like molecules, hopanoids**
  - **stabilize membrane**
  - **found in petroleum**

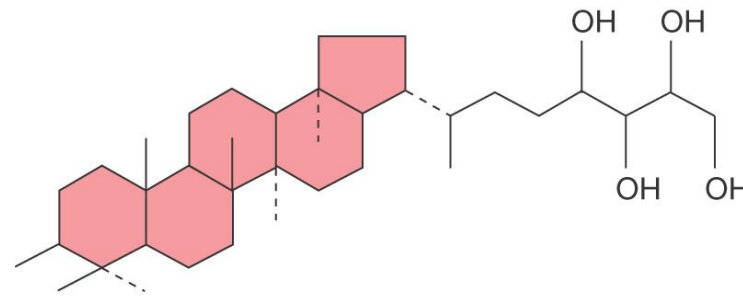


# Figure 3.9 Membrane Steroids and Hopanoids

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(a) Cholesterol (a steroid) is found in eucaryotes



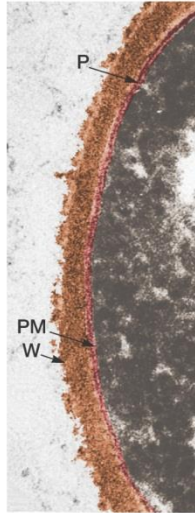
(b) A bacteriohopanetetrol (a hopanoid) is found in bacteria

# Bacterial Cell Wall

- **peptidoglycan (murein)**
  - **rigid structure that lies just outside the cell membrane**
  - **two types based on Gram stain**
    - **gram positive – stain purple; thick peptidoglycan**
    - **gram negative – stain pink or red; thin peptidoglycan and outer membrane**

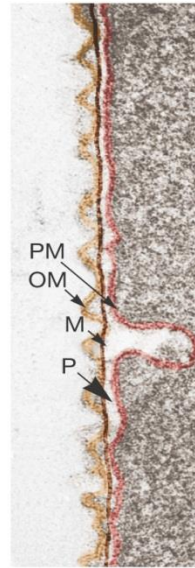
# Figure 3.10

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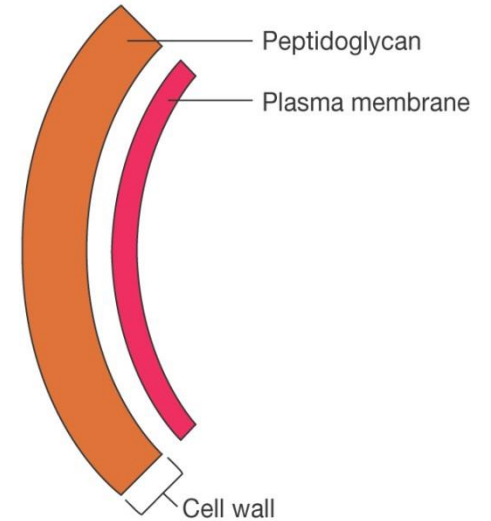
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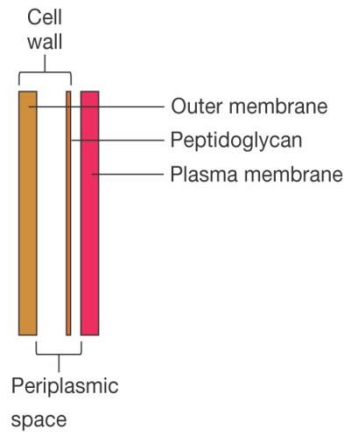
## The gram-positive cell wall



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## The gram-negative cell wall



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# Cell Wall Functions

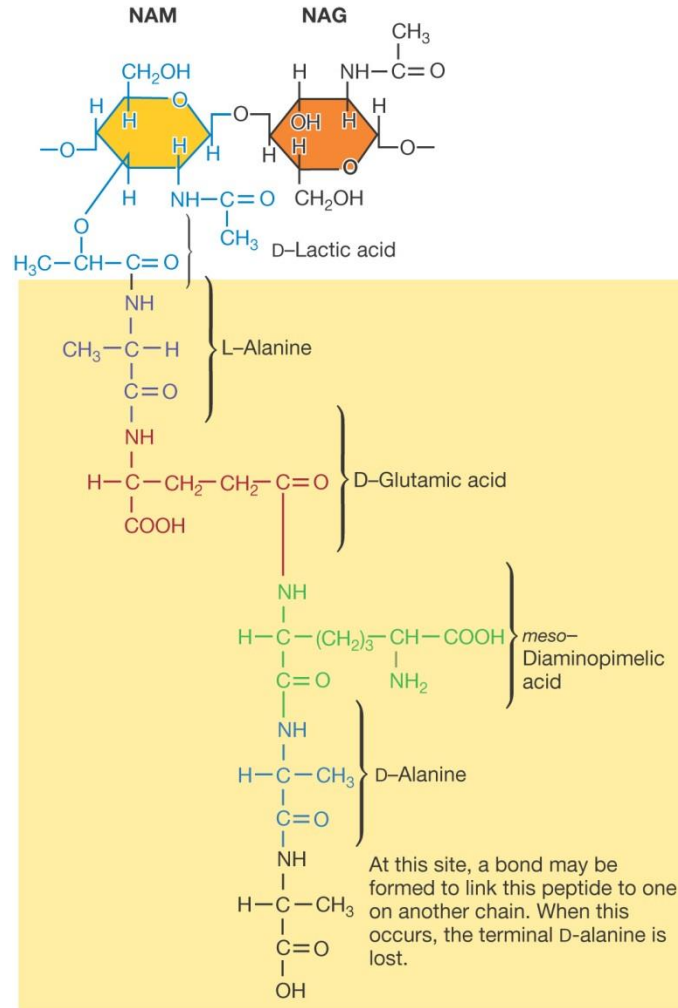
- **maintains shape of the bacterium**
  - **almost all bacteria have one**
- **helps protect cell from osmotic lysis**
- **helps protect from toxic materials**
- **may contribute to pathogenicity**

# Peptidoglycan Structure

- **meshlike polymer of identical subunits forming long strands**
  - **two alternating sugars**
    - ***N*-acetylglucosamine (NAG)**
    - ***N*-acetylmuramic acid**
  - **alternating D- and L- amino acids**

# Figure 3.11

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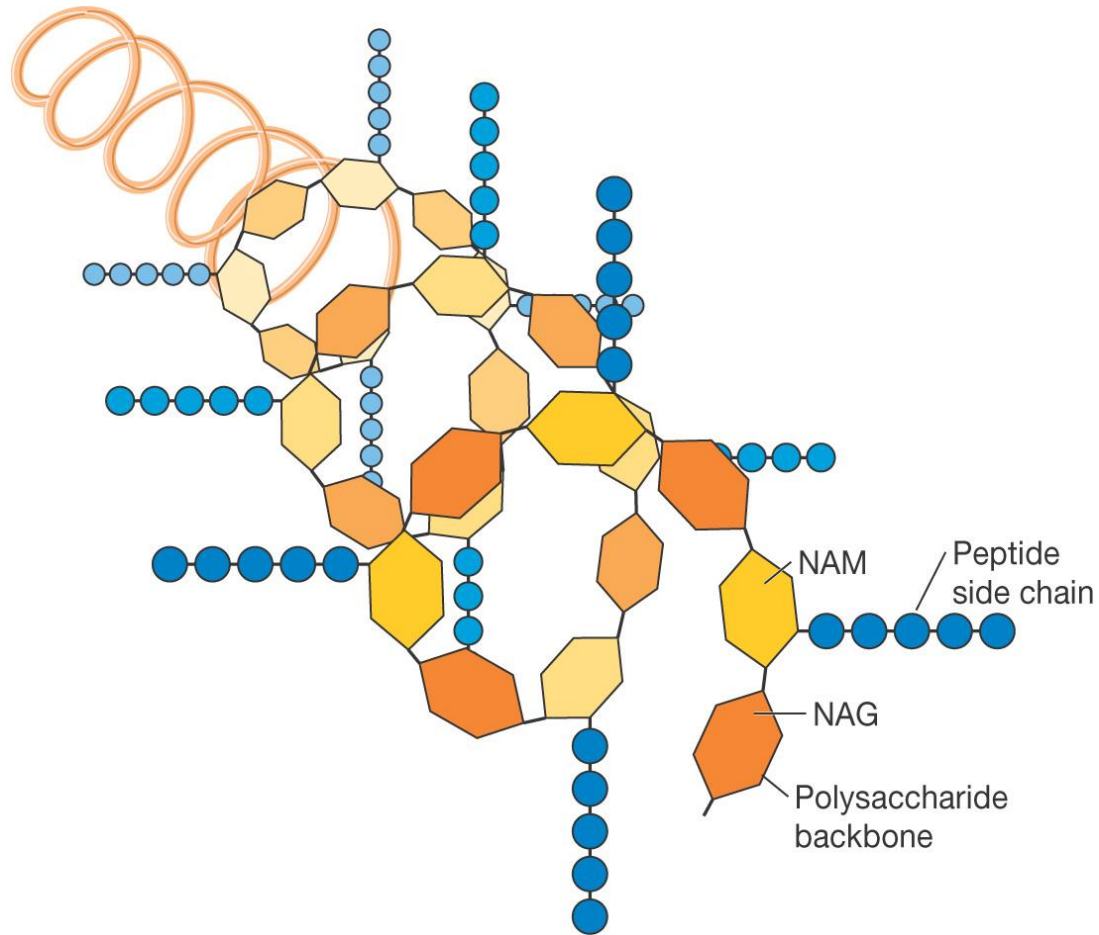


# Strands Are Crosslinked

- **peptidoglycan strands have a helical shape**
- **peptidoglycan chains are crosslinked by peptides for strength**
  - **interbridges may form**
  - **peptidoglycan sacs – interconnected networks**
  - **various structures occur**

# Figure 3.12

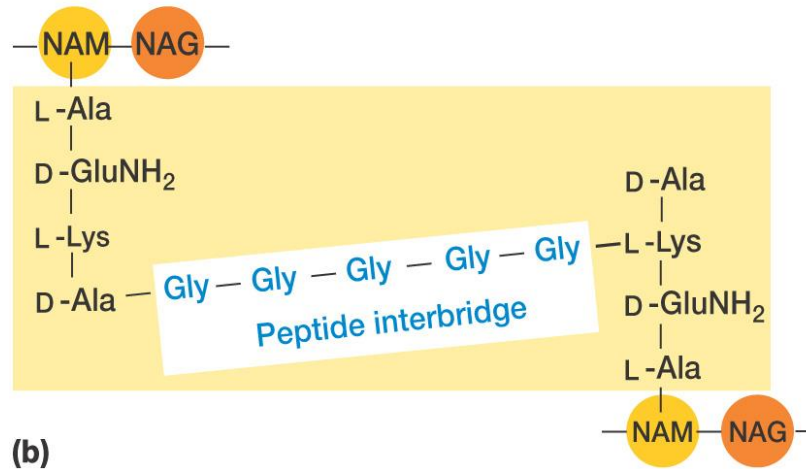
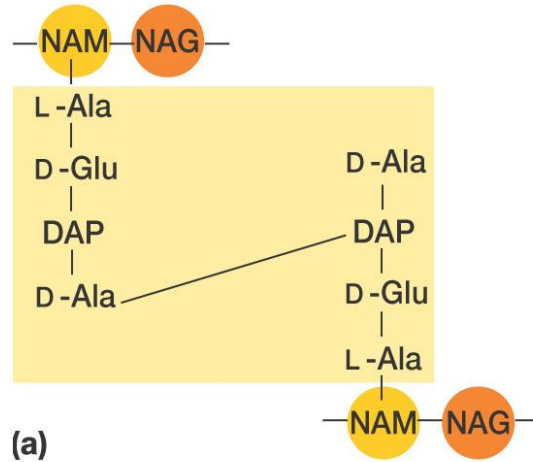
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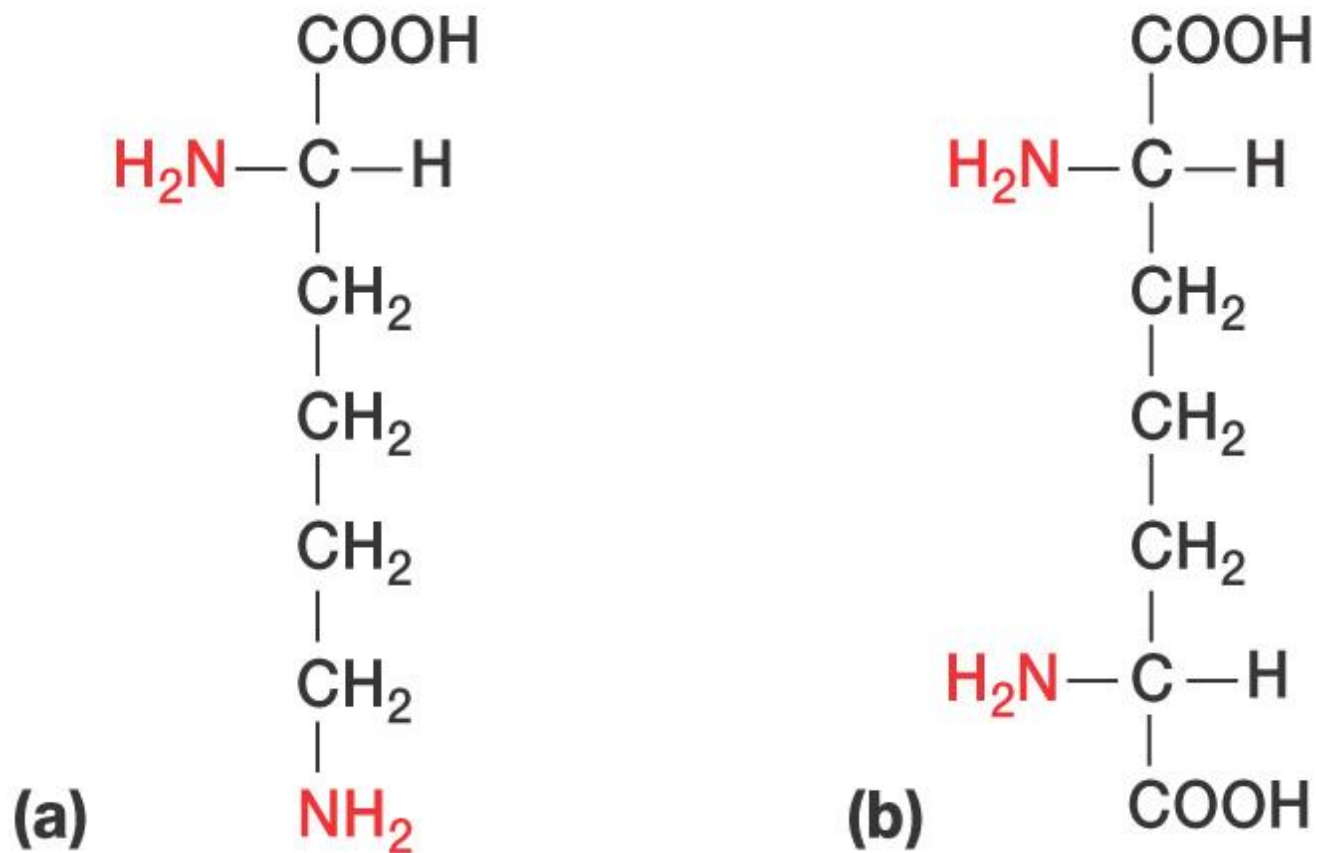
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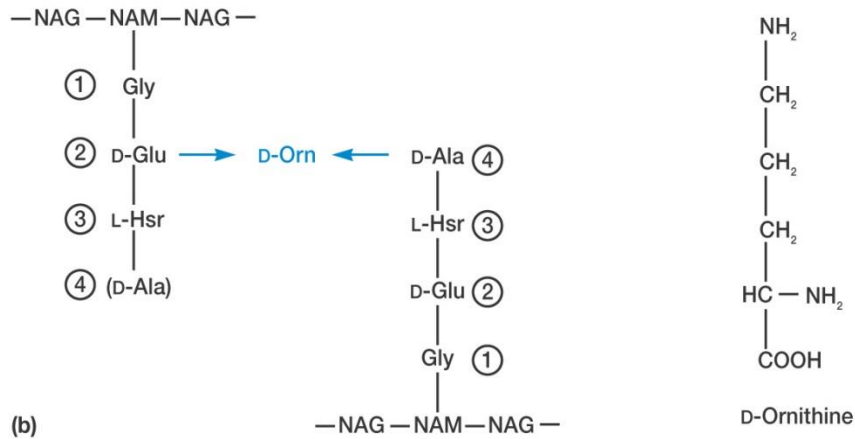
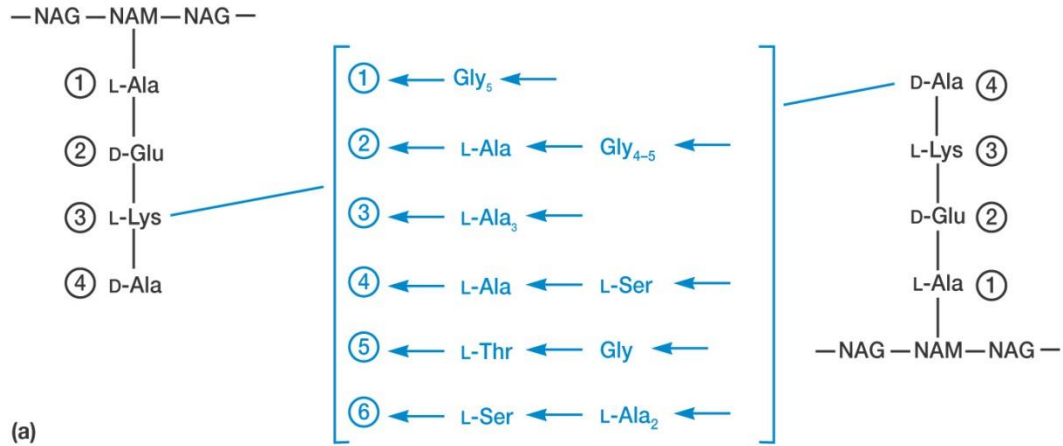
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# Figure 3.16

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# **Gram-Positive Cell Walls**

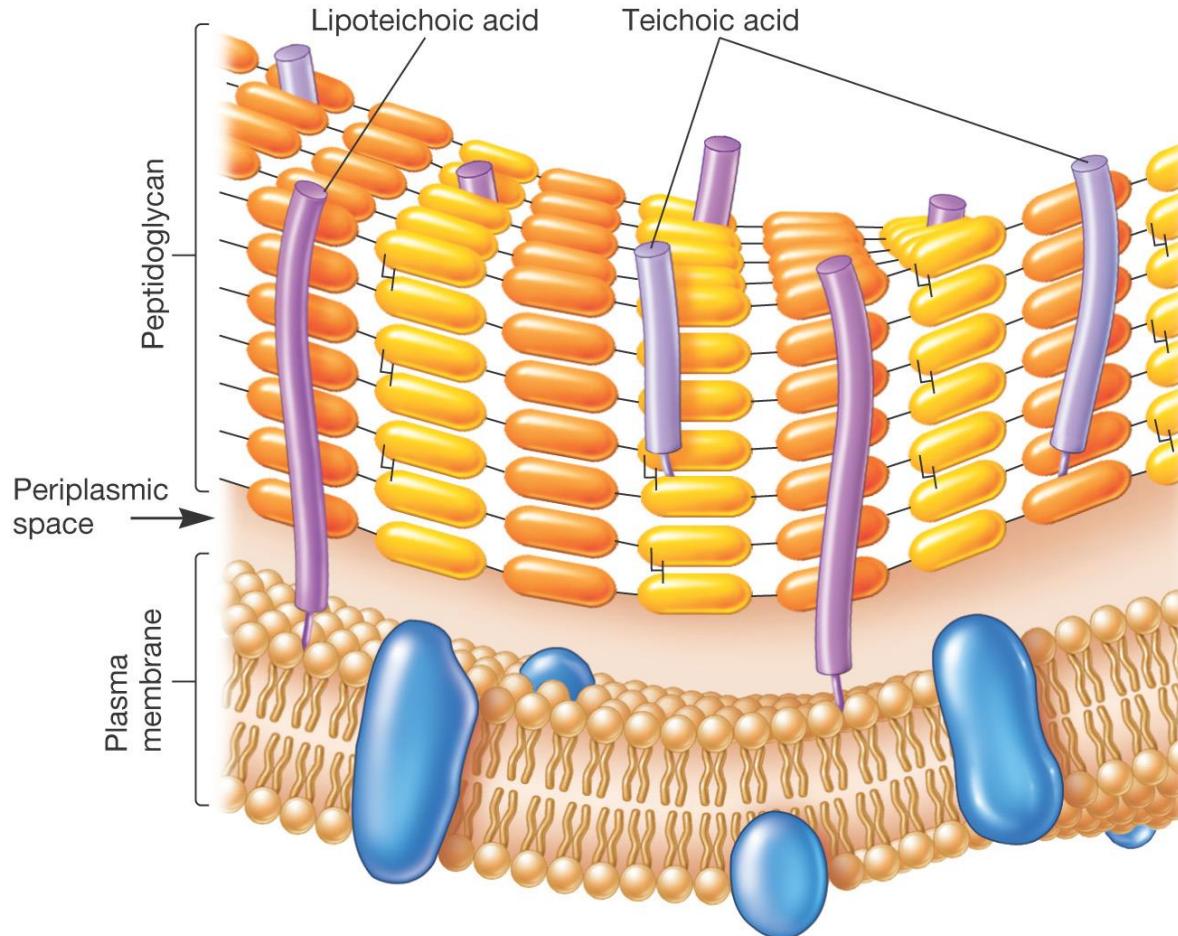
- **composed primarily of peptidoglycan**
- **may also contain large amounts of teichoic acids (negatively charged)**
  - **help maintain cell envelop**
  - **protect from harmful environmental substances**
  - **may bind to host cells**
- **some gram-positive bacteria have layer of proteins on surface of peptidoglycan**

# **Periplasmic Space of Gram +ve Bacteria**

- **lies between plasma membrane and cell wall and is smaller than that of gram-negative bacteria**
- **periplasm has relatively few proteins**
- **enzymes secreted by gram-positive bacteria are called exoenzymes**
  - **aid in degradation of large nutrients**

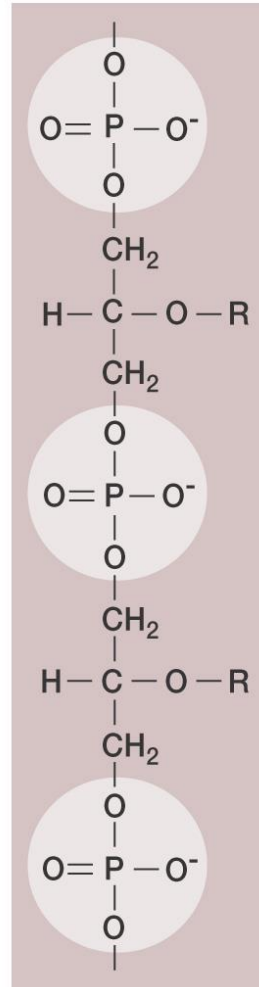
# Figure 3.17; gm+ cell wall

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# Figure 3.18; teichoic acid structure

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# Gram-Negative Cell Walls

- **more complex than gram positive**
- **consist of a thin layer of peptidoglycan surrounded by an outer membrane**
- **outer membrane composed of lipids, lipoproteins, and lipopolysaccharide (LPS)**
- **no teichoic acids**



# Gram-Negative Cell Walls

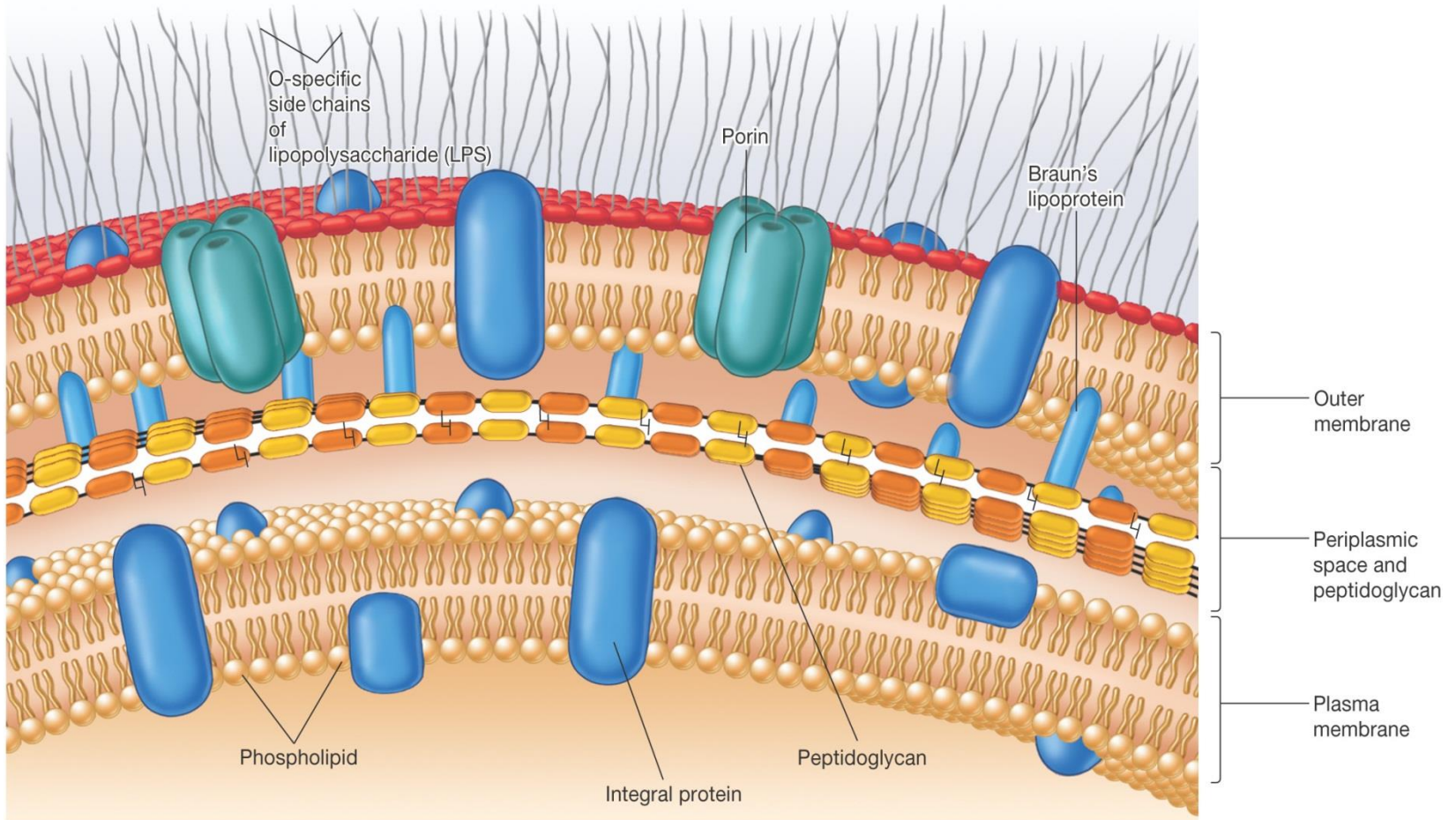
- **peptidoglycan is ~5-10% of cell wall weight**
- **periplasmic space differs from that in gram-positive cells**
  - **may constitute 20–40% of cell volume**
  - **many enzymes present in periplasm**
    - **hydrolytic enzymes, transport proteins and other proteins**

# Gram-Negative Cell Walls

- outer membrane lies outside the thin peptidoglycan layer
- **Braun's lipoproteins** connects outer membrane to peptidoglycan
- other adhesion sites were also reported to be present in the Gm -ve cell wall

# Figure 3.19; typical Gram negative cell wall

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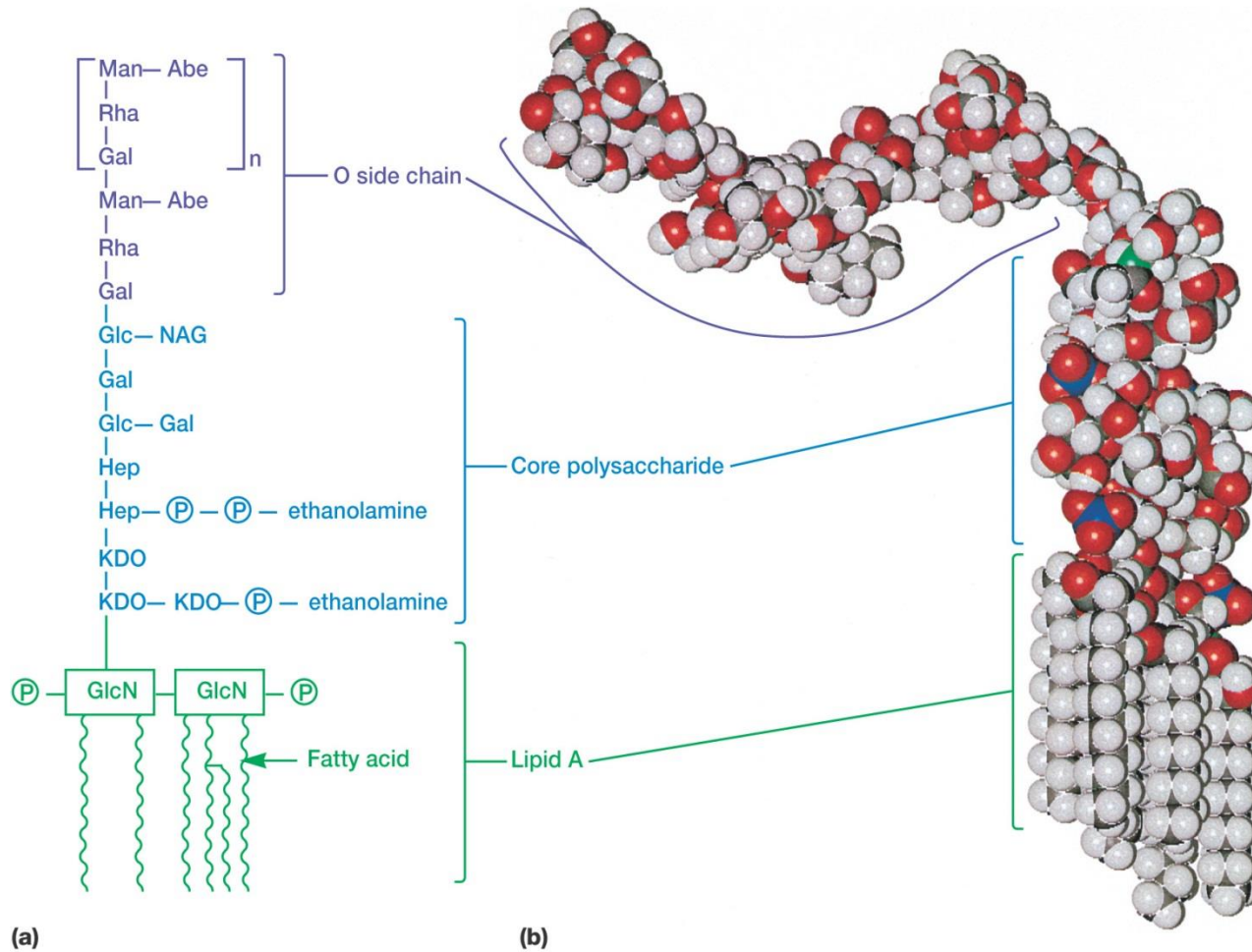


# **Lipopolysaccharides (LPSs)**

- **consists of three parts**
  - **lipid A (toxic)**
  - **core polysaccharide**
  - **O side chain (O antigen); used in bacterial classification and identification**
- **lipid A embedded in outer membrane**
- **core polysaccharide, O side chain extend out from the cell**

# Figure 3.20; LPS

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From M. Kastowsky, T. Gutberlet, and H. Bradaczek, *Journal of Bacteriology*, 774:4798–4806, 1992

# Importance of LPS

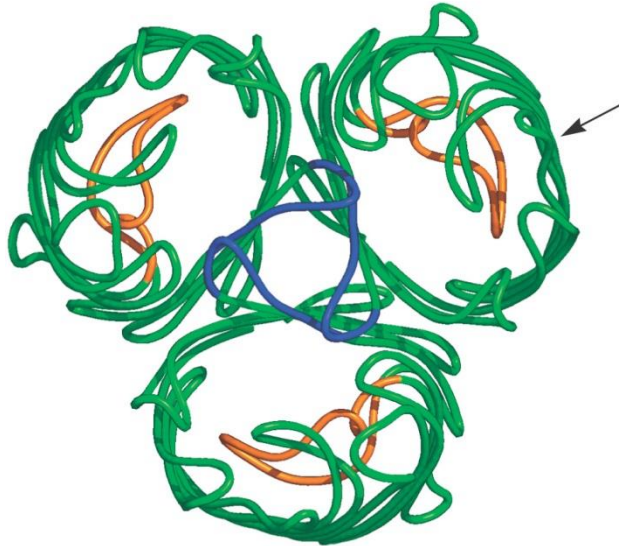
- **contributes to negative charge on cell surface**
- **helps stabilize outer membrane structure**
- **may contribute to attachment to surfaces and biofilm formation**
- **creates a permeability barrier**
- **protection from host defenses (O antigen)**
- **can act as an endotoxin (lipid A)**

# **Gram-Negative Outer Membrane Permeability**

- **more permeable than plasma membrane due to presence of porin proteins and transporter proteins**
  - **porin proteins form channels through which small molecules (600–700 daltons) can pass**

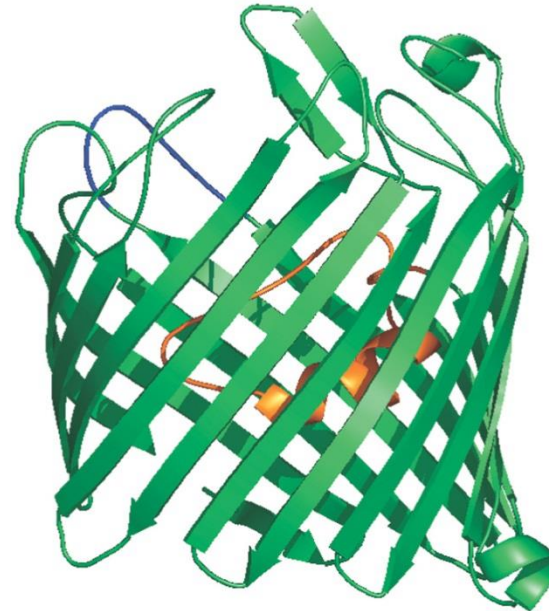
# Figure 3.21; porin proteins

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(a) Porin trimer

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(b) OmpF side view



# Steps of Gram stain

1. fixation of the bacteria on the slide
2. adding gram stain (**violet**)
3. washing
4. adding the counter stain (**red**)

# **Mechanism of Gram Stain Reaction**

- **Gram stain reaction due to nature of cell wall**
- **shrinkage of the pores of peptidoglycan layer of gram-positive cells**
  - **constriction prevents loss of crystal violet during decolorization step**
- **thinner peptidoglycan layer and larger pores of gram-negative bacteria does not prevent loss of crystal violet**

# Cell walls and Osmotic Protection

- **hypotonic environments**
  - solute concentration outside the cell is less than inside the cell
  - water moves into cell and cell swells
  - **cell wall protects from lysis**
- **hypertonic environments**
  - solute concentration outside the cell is greater than inside
  - water leaves the cell
  - **plasmolysis occurs**

# Evidence of Protective Nature of the Cell Wall

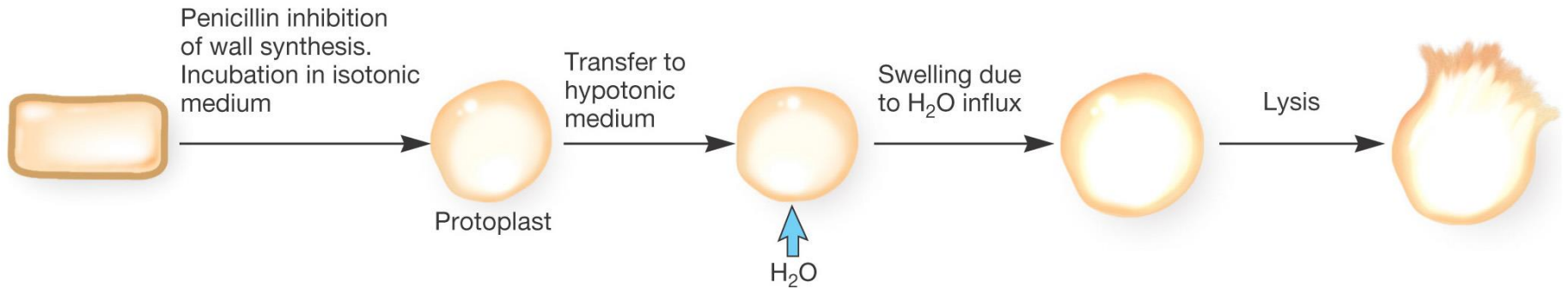
- **lysozyme** breaks the bond between N-acetyl glucosamine and N-acetylmuramic acid
- **penicillin** inhibits peptidoglycan synthesis
- if cells are treated with either of the above they will lyse if they are in a **hypotonic** solution

# Loss of Cell Wall May Survive in Isotonic Environments

- **protoplasts**
- **spheroplasts**
- ***Mycoplasma***
  - **does not produce a cell wall**
  - **Sensitive to osmotic pressure and if placed in hypotonic solution they burst and get destroyed**

# Figure 3.22; protoplast formation and lysis

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# Components Outside of the Cell Wall

- **outermost layer in the cell envelope**
- **Glycocalyx (polysaccharides)**  
**including;**
  - **capsules and slime layers (fuzzy)**
  - **S layers (structured)**
- **aid in attachment to solid surfaces**
  - **e.g., biofilms in plants and animals**

# Capsules

- **usually composed of polysaccharides**
- **well organized and not easily removed from cell**
- **visible in light microscope**
- **protective advantages**
  - **resistant to phagocytosis**
  - **protect from dessication**
  - **exclude viruses and detergents**

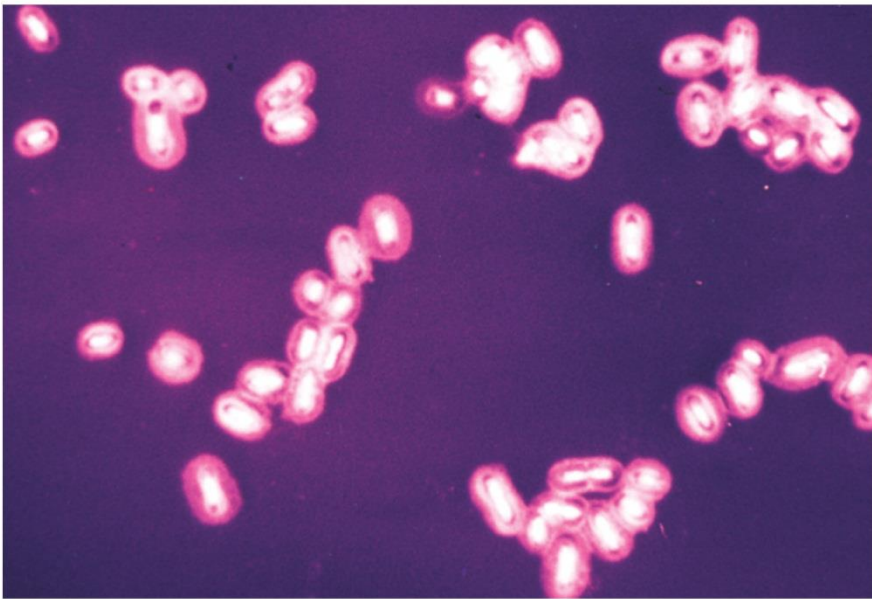


# Slime Layers

- **similar to capsules except diffuse, unorganized and easily removed**
- **slime may aid in motility**

# Figure 3.23; bacterial capsules; *K. pneumoniae*

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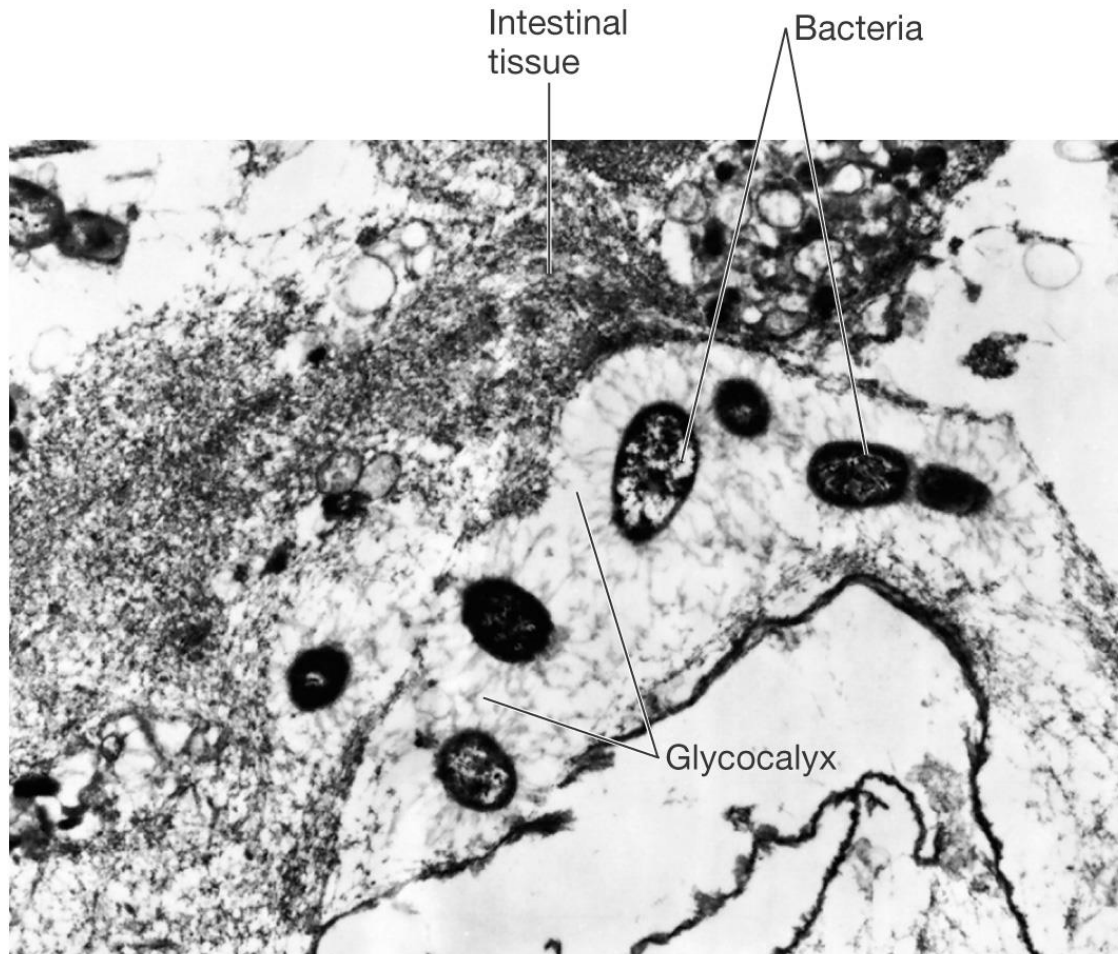
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# Figure 3.24; bacterial glycoalyx

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# **S Layers**

- **regularly structured layers of protein or glycoprotein that self-assemble**
  - **in gram-negative bacteria the S layer adheres to outer membrane**
  - **in gram-positive bacteria it is associated with the peptidoglycan surface**

# S Layer Functions

- **protect from ions and pH fluctuations, osmotic stress, enzymes, and predation**
- **maintains shape and rigidity**
- **promotes adhesion to surfaces**
- **protects from host defenses**
- **potential use in nanotechnology**
  - **S layer spontaneously associates**

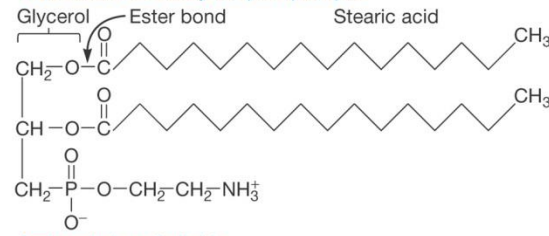
# Archaeal Cell Envelopes

- **differ from bacterial envelopes in the molecular makeup and organization**
  - **S layer may be only component outside plasma membrane**
  - **some lack cell wall**
  - **capsules and slime layers are rare**

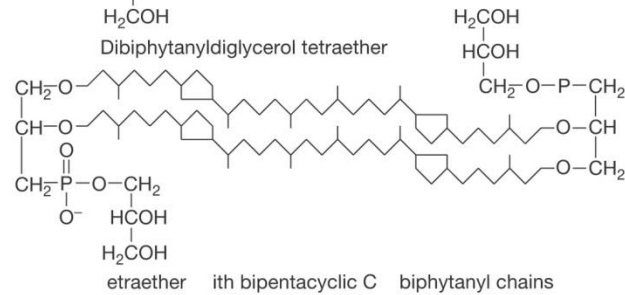
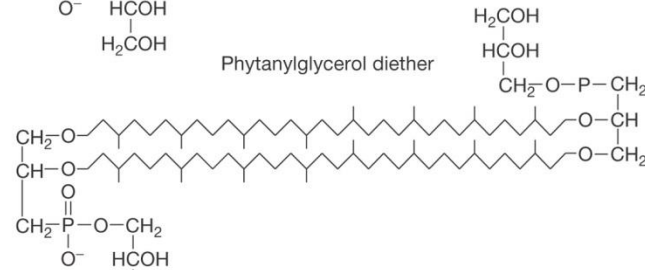
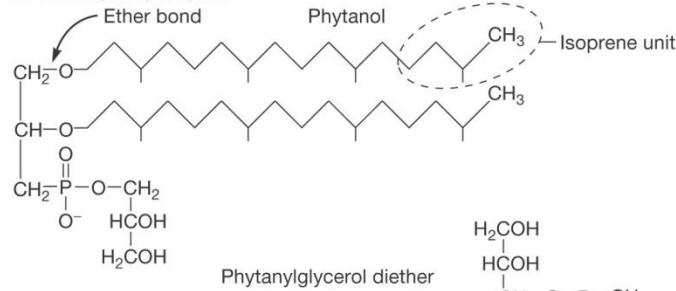
# Archaeal Membranes

- **composed of unique lipids**
  - **isoprene units (five carbon, branched)**
  - **ether linkages rather than ester linkages to glycerol**
- **some have a monolayer structure instead of a bilayer structure**

**A bacterial or eukaryotic phospholipid**



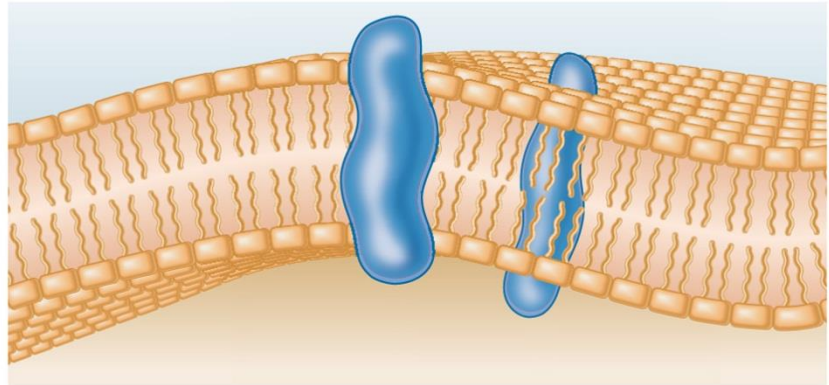
**Archaeal phospholipids**



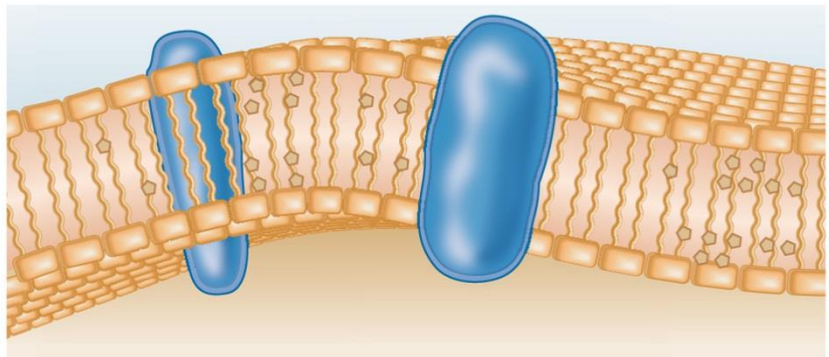


# Figure 3.27

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**(a)** Bilayer of  $C_{20}$  diethers



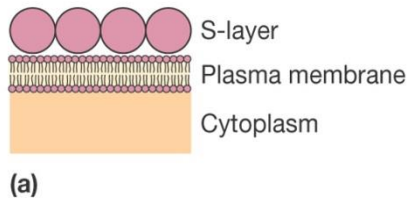
**(b)** Monolayer of  $C_{40}$  tetraethers

# **Archaeal Cell Walls Differ from Bacterial Cell Walls**

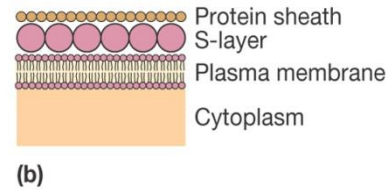
- **lack peptidoglycan**
- **most common cell wall is S layer**
- **may have protein sheath external to S layer**
- **S layer may be outside membrane and separated by pseudomurein**
- **pseudomurein may be outermost layer – similar to gram-positive microorganisms**

# Figure 3.28

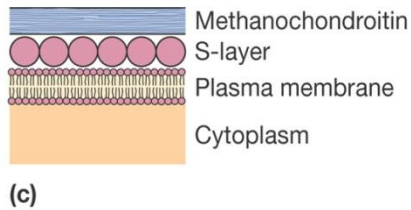
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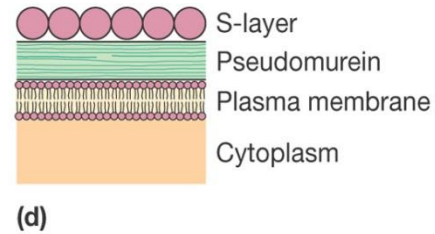
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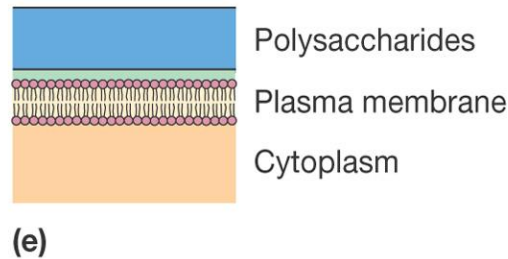
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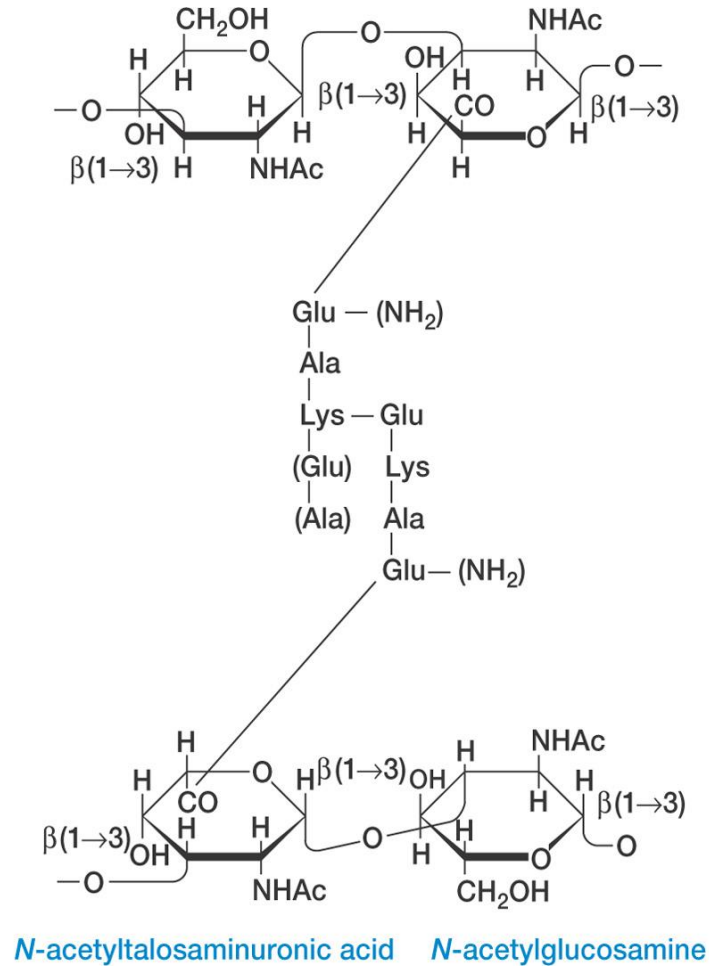


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# Figure 3.29

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# **Bacterial and Archaeal Cytoplasmic Structures**

**Cytoskeleton**

**Intracytoplasmic membranes**

**Inclusions**

**Ribosomes**

**Nucleoid and plasmids**

# Protoplast and Cytoplasm

- **protoplast is plasma membrane and everything within**
- **cytoplasm - material bounded by the plasma membrane**

# The Cytoskeleton

- **homologs of all 3 eukaryotic cytoskeletal elements have been identified in bacteria and 2 in archaea**
- **functions are similar as in eukaryotes**
  - **Role in cell division, protein localization, and determination of cell shape**

# Table 3.2

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**Table 3.2** Bacterial Cytoskeletal Proteins

<i>Type</i>	<i>Function</i>	<i>Comments</i>
<b>Tubulin homologs</b>		
FtsZ	Cell division	Widely observed in <i>Bacteria</i> and <i>Archaea</i>
BtubA/BtubB	Unknown	Observed only in <i>Prosthecoacter</i> ; thought to be encoded by eukaryotic tubulin genes obtained by horizontal gene transfer
<b>Actin homologs</b>		
FtsA	Cell division	Observed in many bacterial species
MamK	Positioning magnetosomes	Observed in magnetotactic species
MreB/Mbl	Maintains cell shape, segregates chromosomes, localizes proteins	Most rod-shaped bacteria
<b>Intermediate filament homologs</b>		
CreS (crescentin)	Induces curvature in curved rods	<i>Caulobacter crescentus</i>
<b>Unique bacterial cytoskeletal proteins</b>		
MinD	Prevents polymerization of FtsZ at cell poles	Many rod-shaped bacteria
ParA (chromosome-encoded form)	Segregates chromosomes	Observed in many species including <i>Vibrio cholerae</i> and <i>C. crescentus</i>

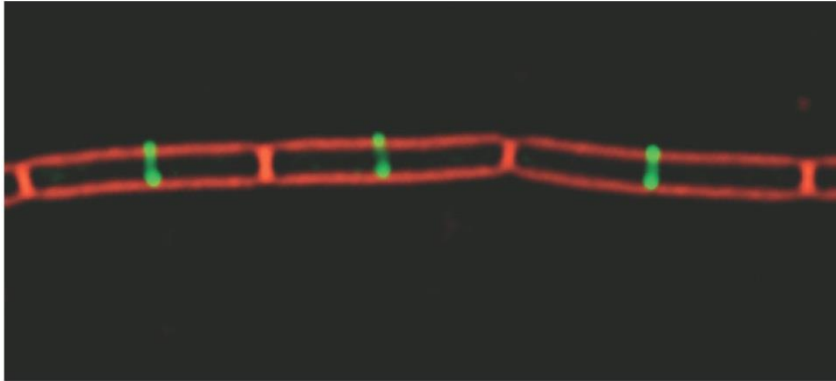


# Best Studied Examples

- **FtsZ** – many bacteria and archaea
  - forms ring during septum formation in cell division
- **MreB** – many rods, some archaea
  - maintains shape by positioning peptidoglycan synthesis machinery
- **CreS** – rare, maintains curve shape

## Figure 3.30; Ftsz, mbl and crescentin

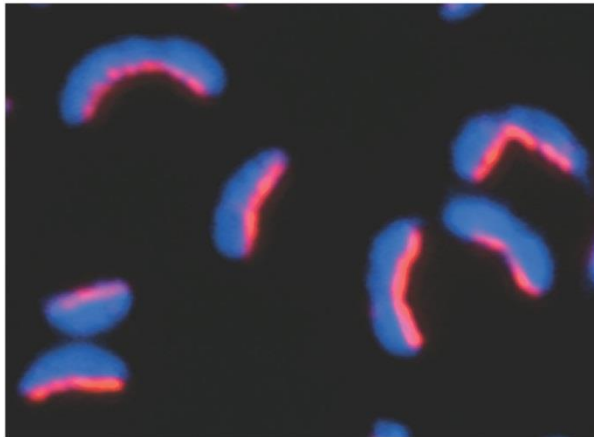
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(a) FtsZ

Dr. Joseph Pogliano

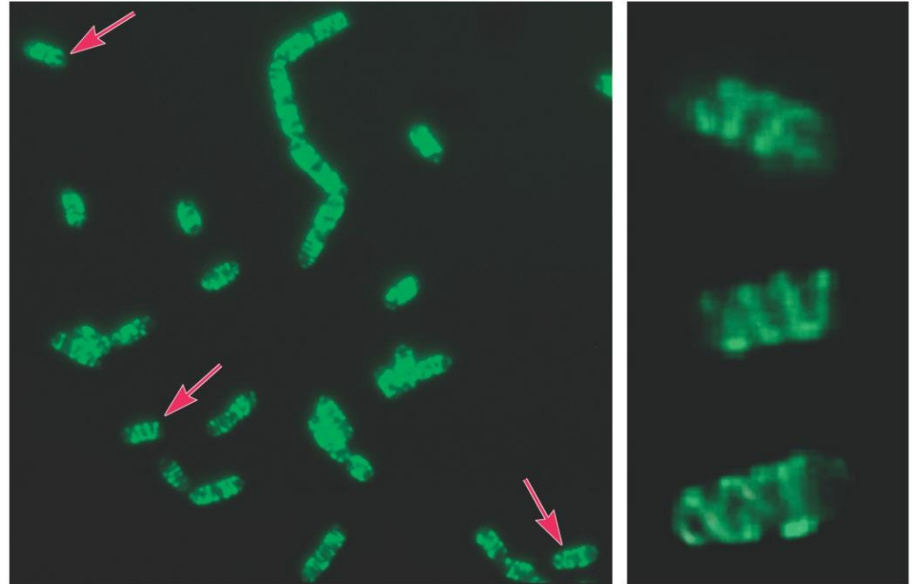
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(d) Crescentin

Dr. Chistine Jacobs-Wagner

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(b) Mbl

(c) Mbl

Image courtesy of Rut Carballido-Lo'pez and Jeff Errington

# Intracytoplasmic Membranes

- **plasma membrane infoldings**
  - **observed in many photosynthetic bacteria**
    - **analogous to thylakoids of chloroplasts**
    - **reactions centers for ATP formation**
  - **observed in many bacteria with high respiratory activity**
- **anammoxosome in *Planctomyces***
  - **organelle – site of anaerobic ammonia oxidation**

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# Figure 3.3; internal bacterial membranes

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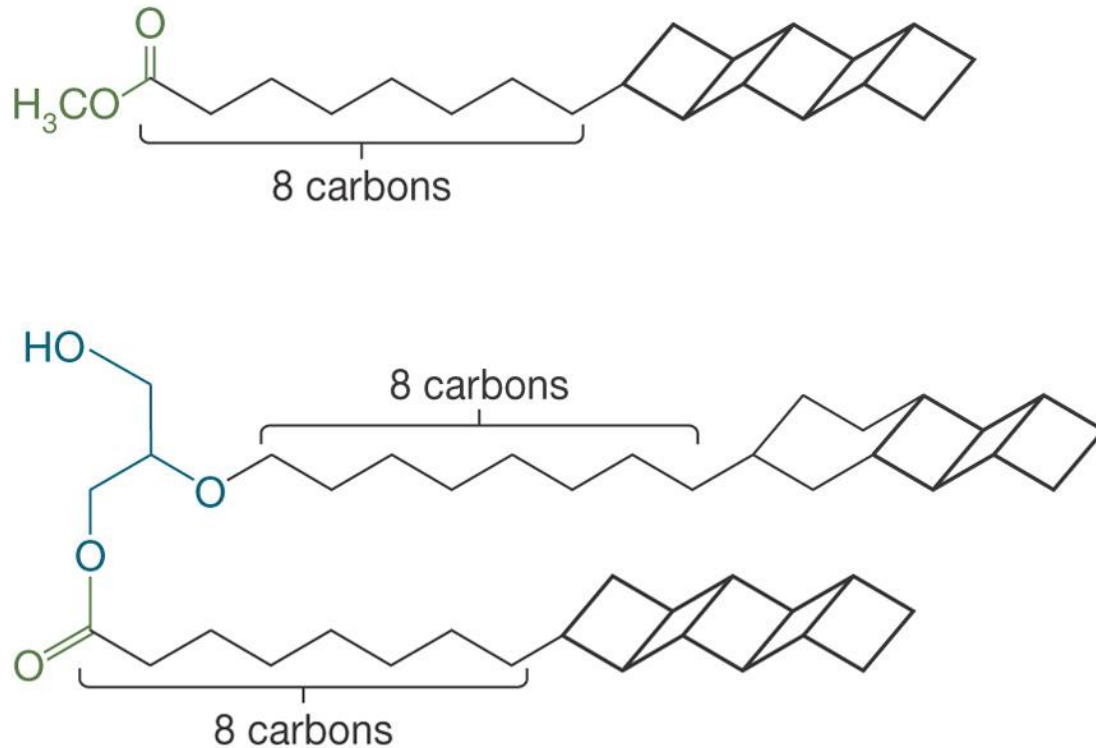
American Society for Microbiology



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## Figure 3.32; ladderane lipids; these are unique to planctomycetes

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# Inclusions

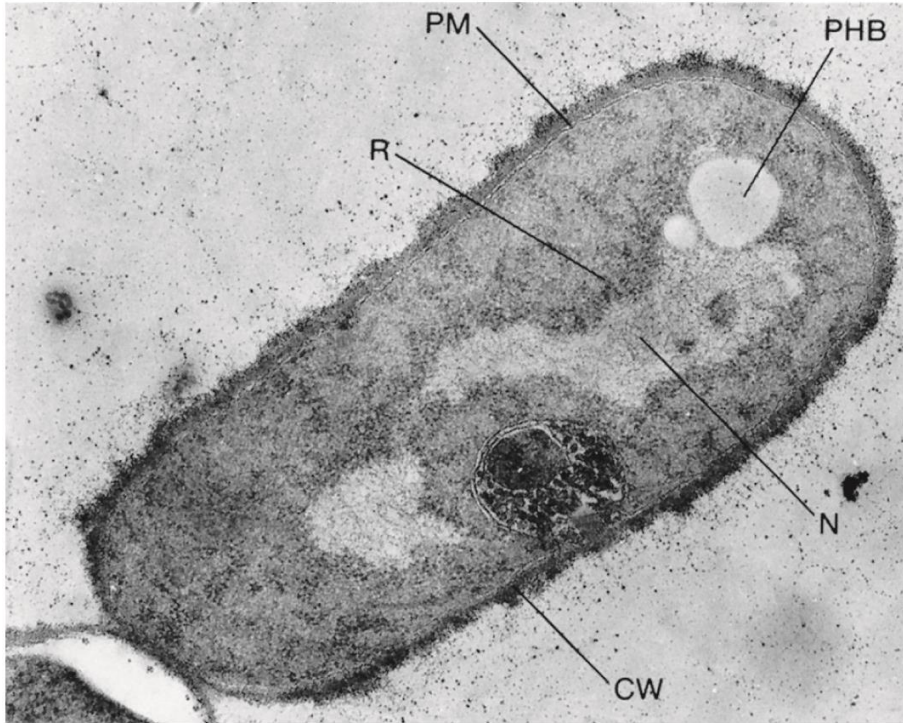
- **granules of organic or inorganic material that are stockpiled by the cell for future use**
- **some are enclosed by a single-layered membrane**
  - **membranes vary in composition**
  - **some made of proteins; others contain lipids**
  - **may be referred to as microcompartments**

# Storage Inclusions

- **storage of nutrients, metabolic end products, energy, building blocks**
- **glycogen storage**
- **carbon storage**
  - **poly- $\beta$ -hydroxybutyrate (PHB)**
- **phosphate - Polyphosphate (Volutin)**
- **amino acids - cyanophycin granules**

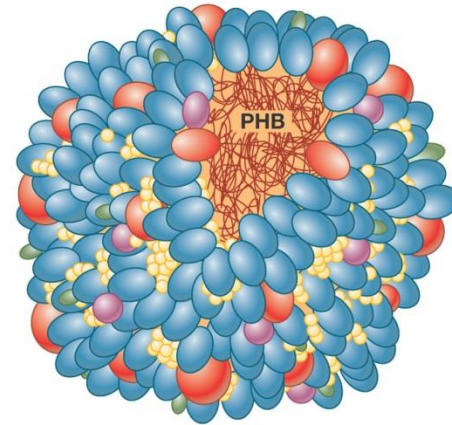
# Figure 3.33

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(b)



# Figure 3.34

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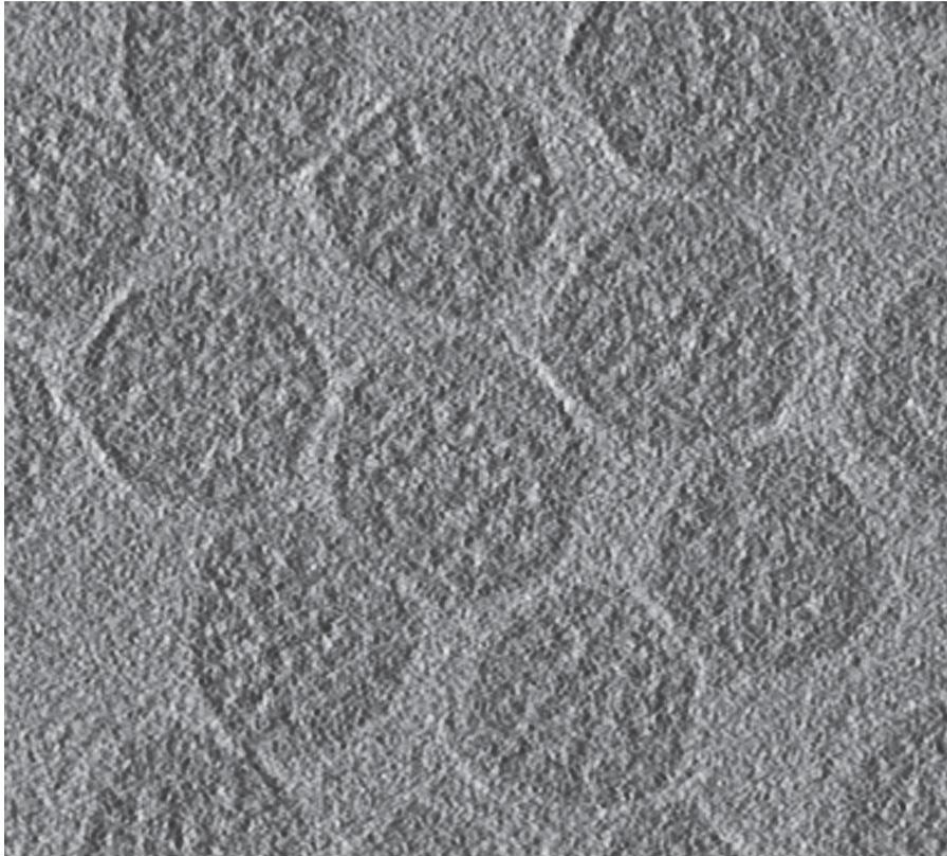
Reprinted from The Shorter Bergey's Manual of Determinative Bacteriology,  
8e, John G. Holt, Editor, 1977 © Bergey's Manual Trust.  
Published by Williams & Wilkins Baltimore, MD;

# Microcompartments

- **not bound by membranes but compartmentalized for a specific function**
- **carboxysomes - CO<sub>2</sub> fixing bacteria**
  - **contain the enzyme ribulose-1,5,-bisphosphate carboxylase (Rubisco), enzyme used for CO<sub>2</sub> fixation**

# Figure 3.35

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Michael Schmid

# Other Inclusions

- **gas vacuoles**
  - found in aquatic, photosynthetic bacteria and archaea
  - provide buoyancy in gas vesicles
- **magnetosomes**
  - found in aquatic bacteria
  - magnetite particles for orientation in Earth's magnetic field
  - cytoskeletal protein MamK
    - helps form magnetosome chain

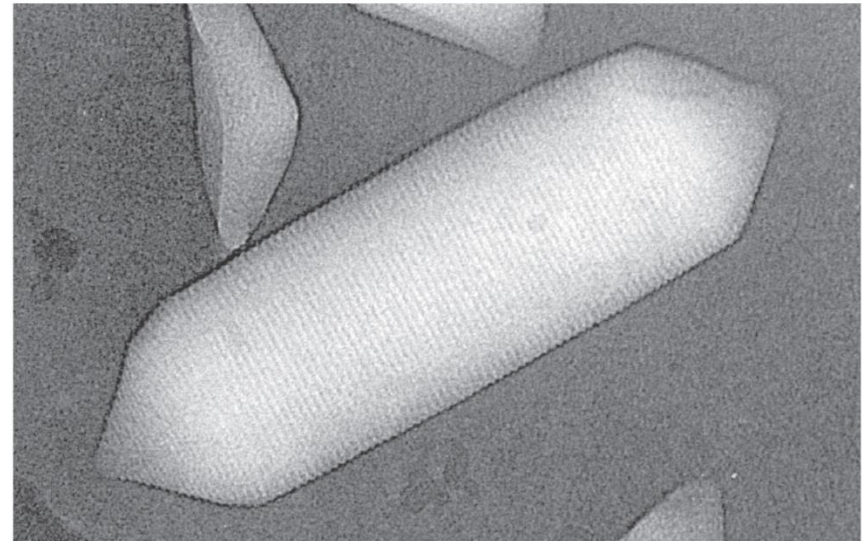
# Figure 3.36

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Courtesy of Daniel Branton, Harvard University

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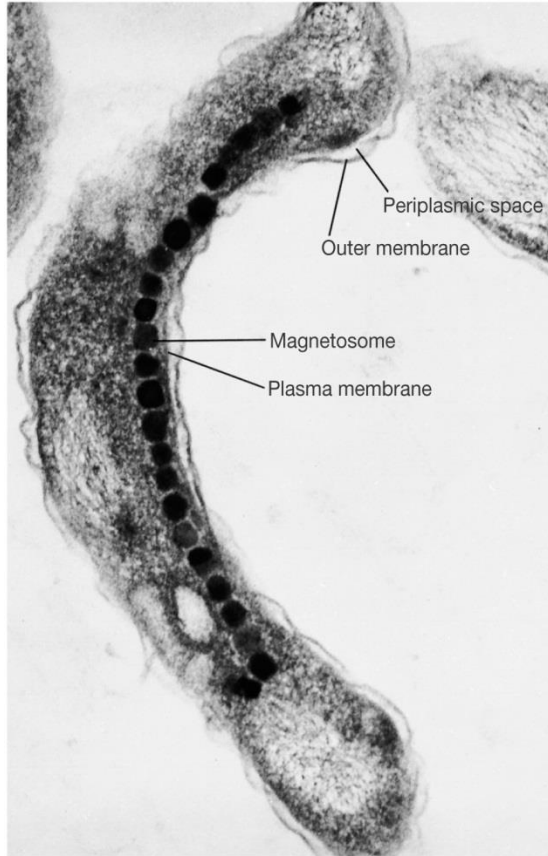
(b)

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# Figure 3.37

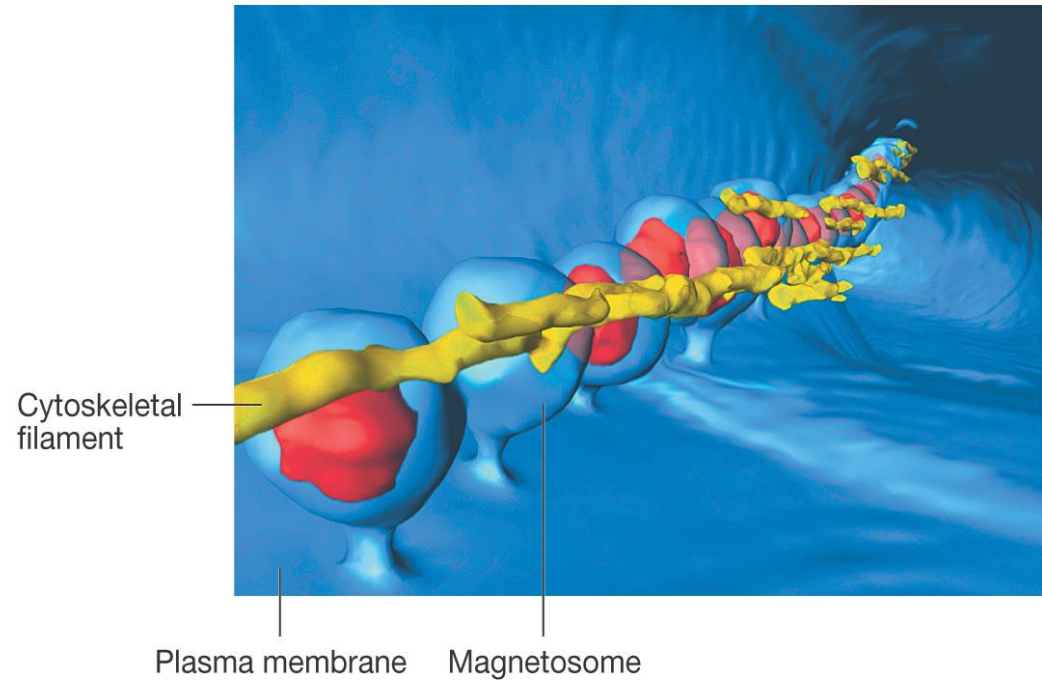
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(a)

Y. Gorby

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(b)

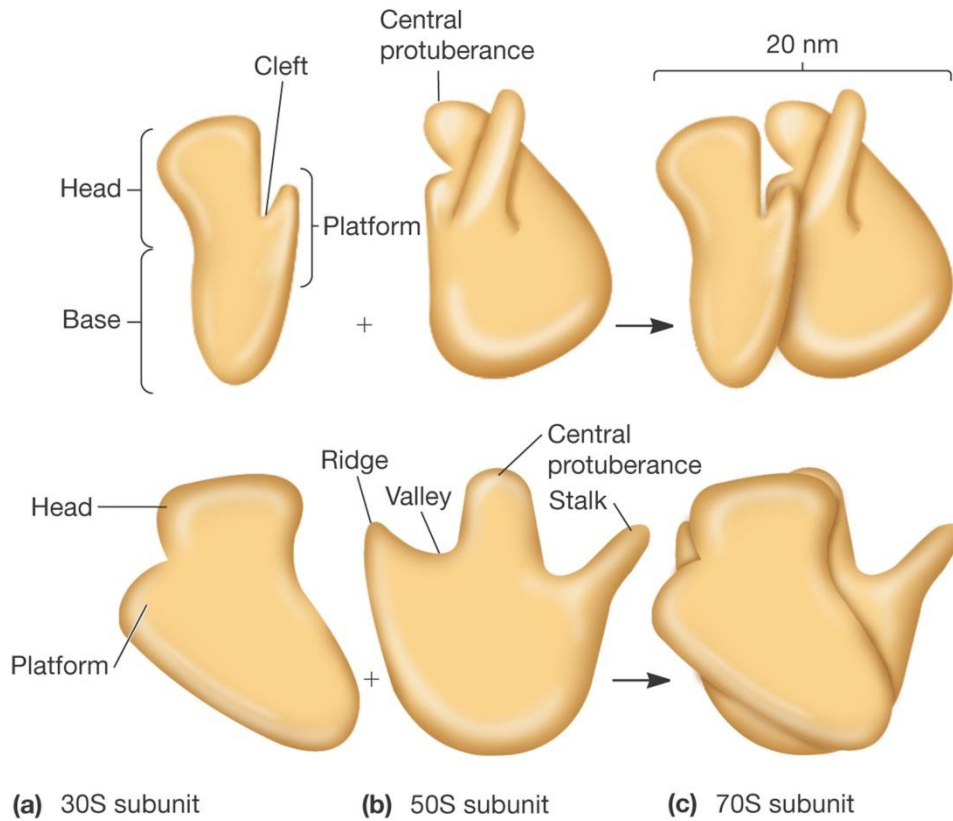
Zhuo Li and Grant Jensen

# Ribosomes

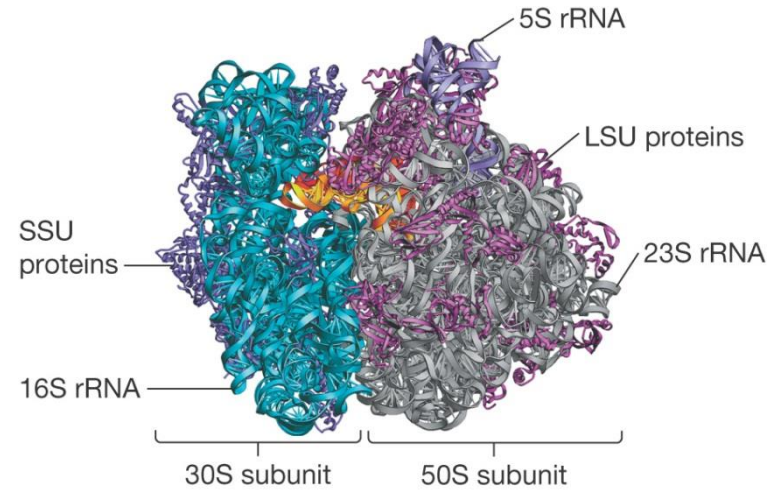
- **complex structures**
  - consisting of protein and RNA
  - sites of protein synthesis
- **entire ribosome**
  - bacterial and archaea ribosome = 70S
  - eukaryotic (80S) S = Svedburg unit
- **bacterial and archaeal ribosomal RNA**
  - 16S small subunit
  - 23S and 5S in large subunit
  - archaea has additional 5.8S (also seen in eukaryotic large subunit)
- **proteins vary**
  - archaea more similar to eukarya than to bacteria

# Figure 3.38

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Harry Noller, University of California, Santa Cruz

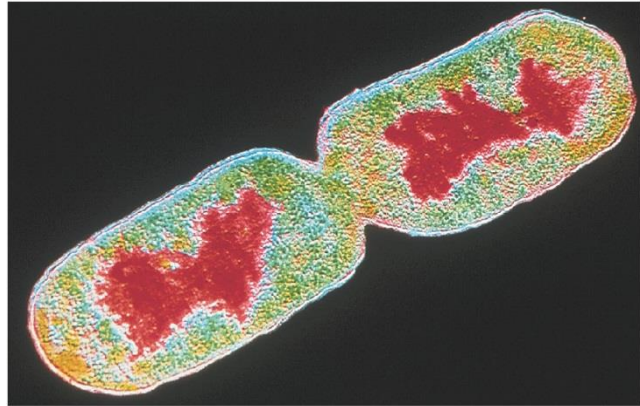


# The Nucleoid

- **irregularly shaped region in bacteria and archaea**
- **usually not membrane bound (few exceptions)**
- **location of chromosome and associated proteins**
- **usually 1**
  - **a closed circular, double-stranded DNA molecule**
- **supercoiling and nucleoid proteins (HU) probably aid in folding**
  - **nucleoid proteins differ from histones**

# Figure 3.39

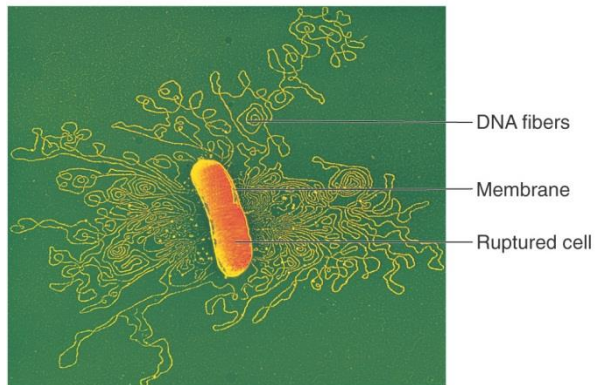
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(a)

0.5  $\mu\text{m}$

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(b)

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(c)

500 nm

Ohniwa R, Morikawa K, Kim J, Kobori T, Hizume K, et al. 2007. *Microsec. Microanal.* 13:3–12

# Plasmids

- **extrachromosomal DNA**
  - found in bacteria, archaea, some fungi
  - usually small, closed circular DNA molecules
- **exist and replicate independently of chromosome**
  - episomes – may integrate into chromosome
- **contain few genes that are non-essential**
  - confer selective advantage to host (e.g., drug resistance)

# Plasmids

- **may exist in many copies in cell**
- **inherited stably during cell division**
- **curing is the loss of a plasmid**
- **classification of plasmids based on mode of existence, spread, and function**
- **see Table 3.3**

# Table 3.3

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<b>Table 3.3 Major Types of Bacterial Plasmids</b>					
<i>Type</i>	<i>Representatives</i>	<i>Approximate Size (kbp)</i>	<i>Copy Number (Copies/Chromosome)</i>	<i>Hosts</i>	<i>Phenotypic Features<sup>a</sup></i>
<b>Conjugative Plasmids<sup>b</sup></b>	F factor	95–100	1–3	<i>E. coli</i> , <i>Salmonella</i> , <i>Citrobacter</i>	Sex pilus, conjugation
<b>R Plasmids</b>	RP4	54	1–3	<i>Pseudomonas</i> and many other gram-negative bacteria	Sex pilus, conjugation, resistance to Amp, Km, Nm, Tet
	pSH6	21		<i>Staphylococcus aureus</i>	Resistance to Gm, Tet, Km
<b>Col Plasmids</b>	ColE1	9	10–30	<i>E. coli</i>	Colicin E1 production
	CloDF13	10	50–70	<i>E. coli</i>	Cloacin DF13
<b>Virulence Plasmids</b>	Ent (P307)	83		<i>E. coli</i>	Enterotoxin production
	Ti	200		<i>Agrobacterium tumefaciens</i>	Tumor induction in plants
<b>Metabolic Plasmids</b>	CAM	230		<i>Pseudomonas</i>	Camphor degradation
	TOL	75		<i>Pseudomonas putida</i>	Toluene degradation

<sup>a</sup>Abbreviations used for resistance to antibiotics: Amp, ampicillin; Gm, gentamycin; Km, kanamycin; Nm, neomycin; Tet, tetracycline.<sup>b</sup>Many R plasmids, metabolic plasmids, and others are also conjugative.

# External Structures

- **extend beyond the cell envelop in bacteria and archaea**
- **function**
  - **protection, attachment to surfaces, horizontal gene transfer, cell movement**
- **pili and fimbriae**
- **flagella**

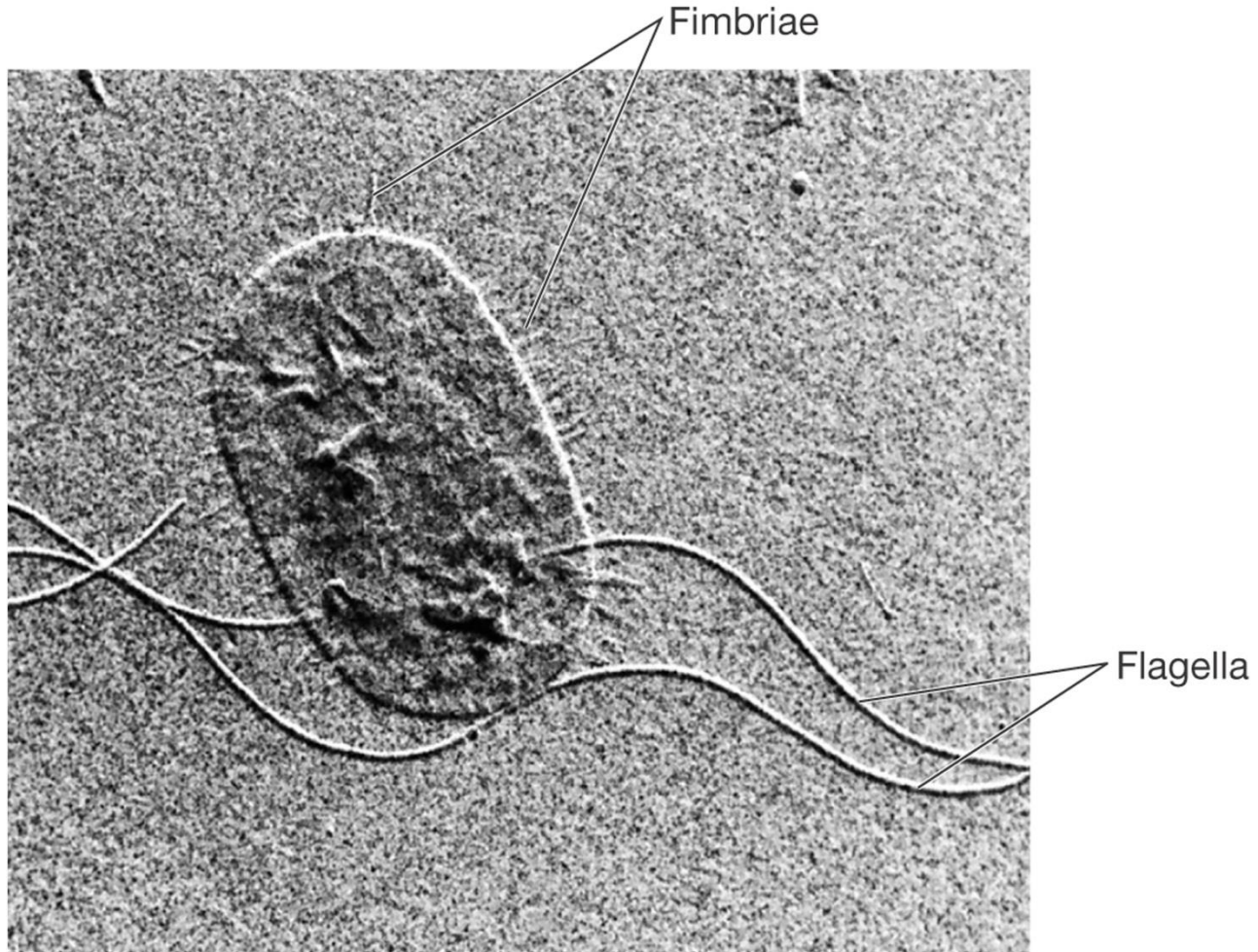
# Pili and Fimbriae

- **fimbriae (s., fimbria); pili (s., pilus)**
  - short, thin, hairlike, proteinaceous appendages (up to 1,000/cell)
  - mediate attachment to surfaces
  - some (type IV fimbriae) required for motility or DNA uptake
- **sex pili (s., pilus)**
  - similar to fimbriae except longer, thicker, and less numerous (1-10/cell)
  - genes for formation found on plasmids
  - required for conjugation



# Figure 3.40

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# Flagella

- **threadlike, locomotor appendages extending outward from plasma membrane and cell wall**
- **functions**
  - **motility and swarming behavior**
  - **attachment to surfaces**
  - **may be virulence factors**

# Bacterial Flagella

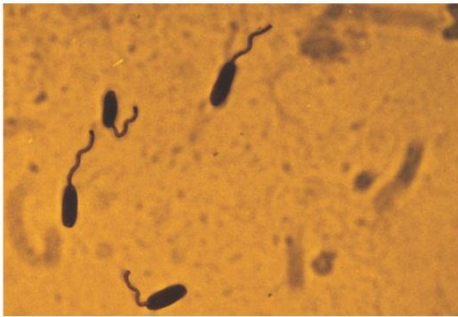
- **thin, rigid protein structures that cannot be observed with bright-field microscope unless specially stained**
- **ultrastructure composed of three parts**
- **pattern of flagellation varies**

# Patterns of Flagella Distribution

- **monotrichous – one flagellum**
- **polar flagellum – flagellum at end of cell**
- **amphitrichous – one flagellum at each end of cell**
- **lophotrichous – cluster of flagella at one or both ends**
- **peritrichous – spread over entire surface of cell**

# Figure 3.41

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(a) *Pseudomonas*—monotrichous polar flagellation

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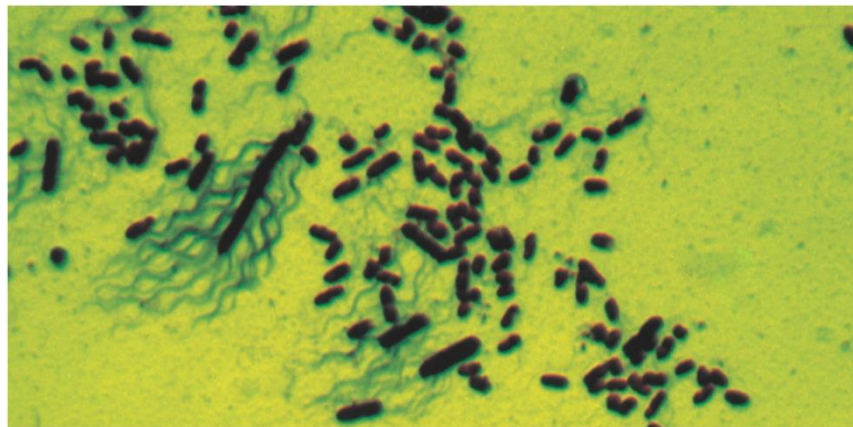
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(b) *Spirillum*—lophotrichous flagellation

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(c) *P. vulgaris*—peritrichous flagellation

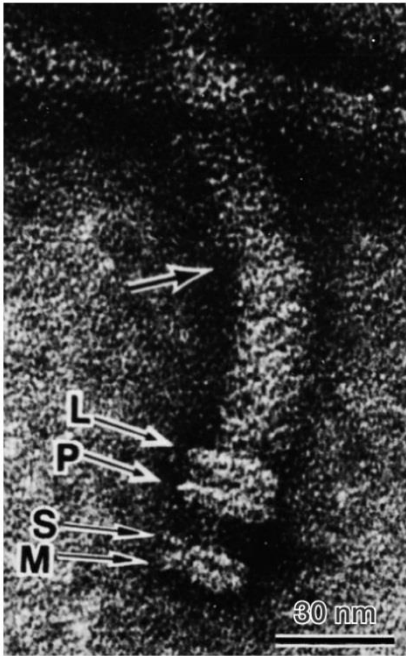
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# Three Parts of Flagella

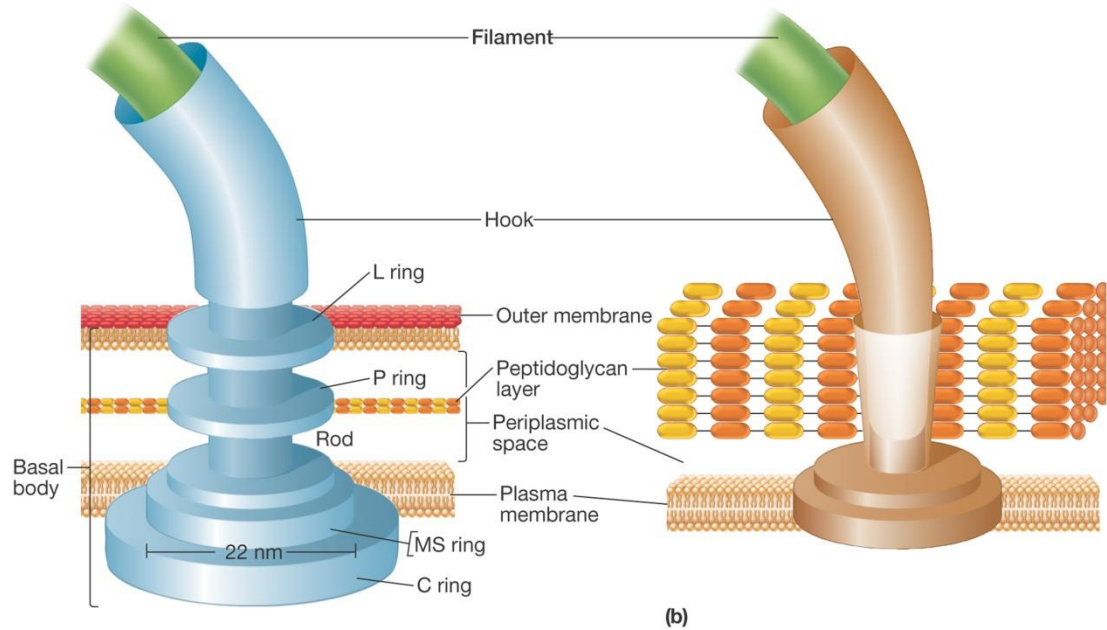
- **filament**
  - extends from cell surface to the tip
  - hollow, rigid cylinder
  - composed of the protein flagellin
  - some bacteria have a sheath around filament
- **hook**
  - links filament to basal body
- **basal body**
  - series of rings that drive flagellar motor

# Figure 3.42

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(a)

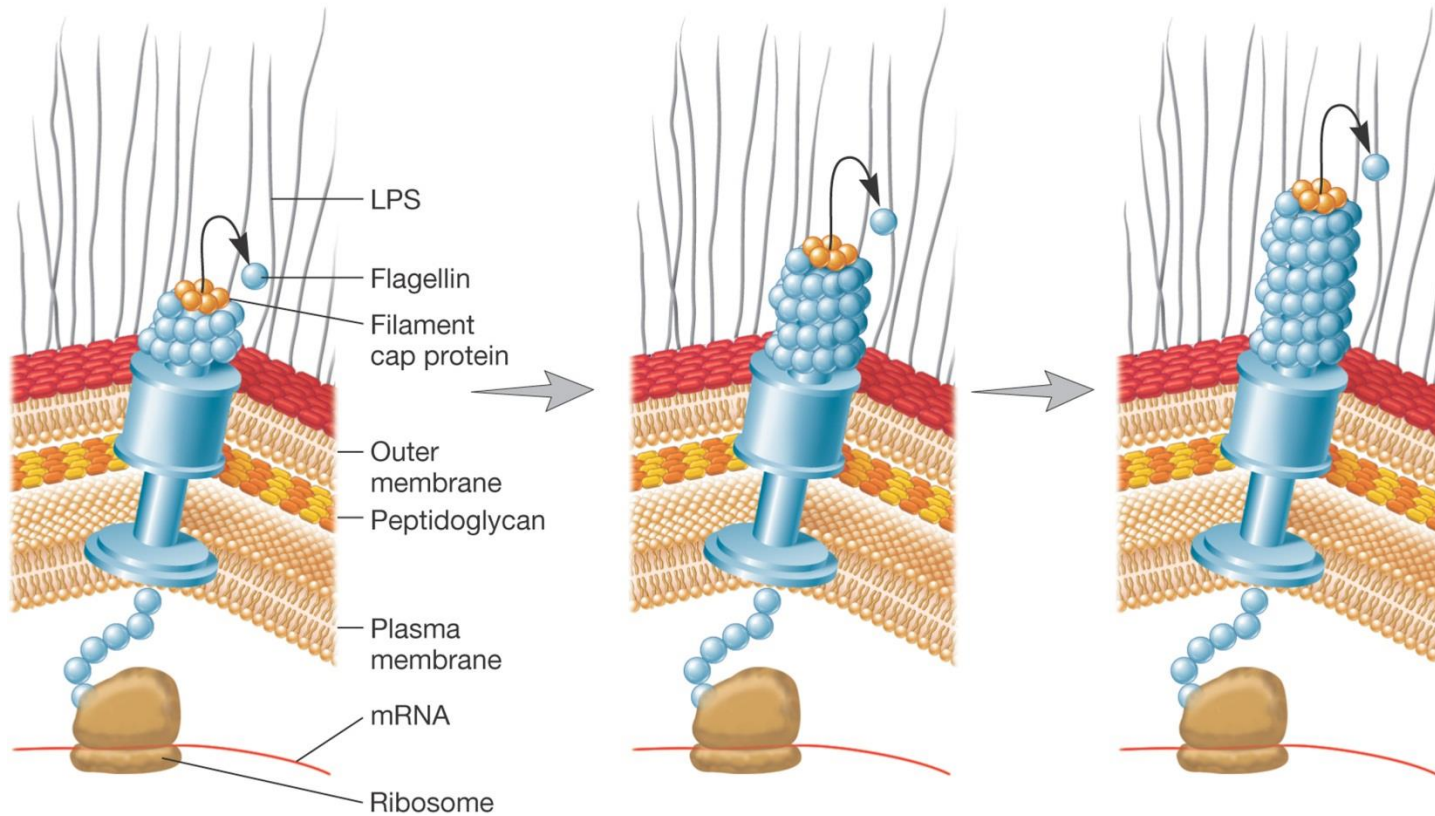
Courtesy of Dr. Julius Adler

# Flagellar Synthesis

- **complex process involving many genes and gene products**
- **new molecules of flagellin are transported through the hollow filament using Type III-like secretion system**
- **filament subunits self-assemble with help of filament cap**
- **growth is from tip, not base**

# Figure 3.43

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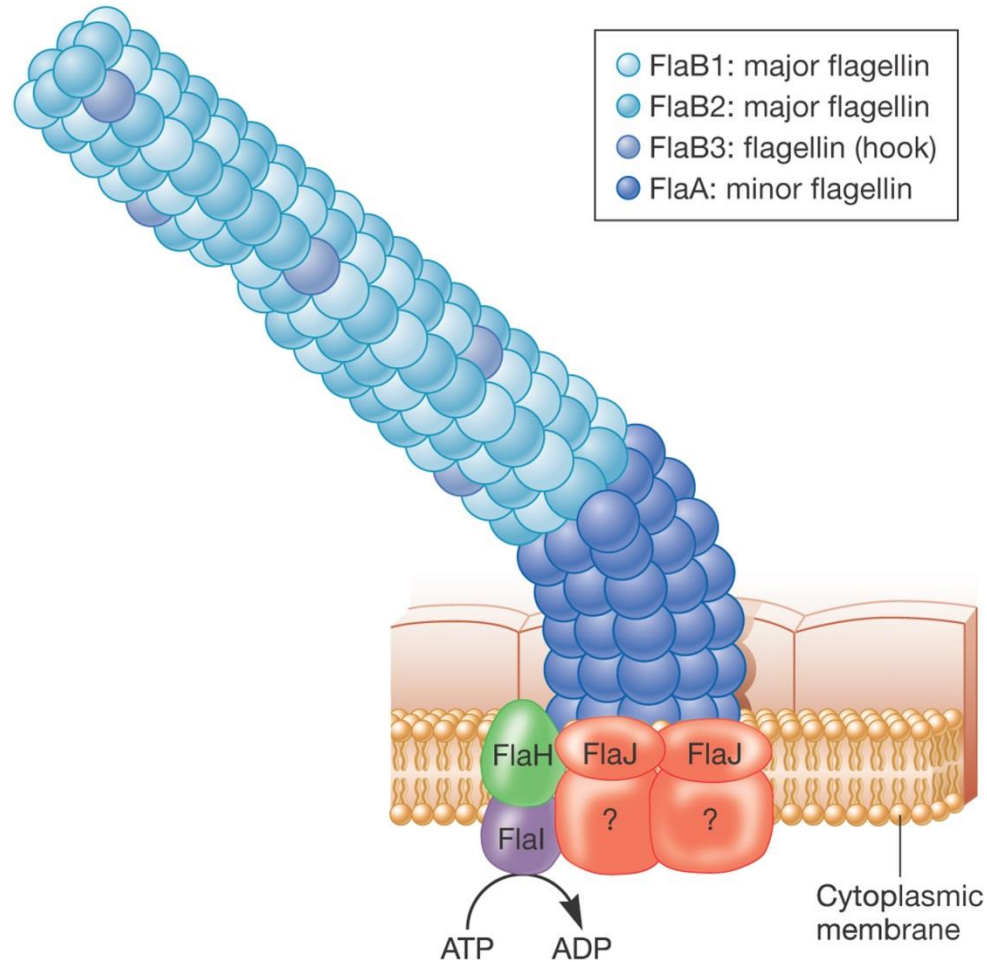


# Differences of Archaeal Flagella

- **flagella thinner**
- **more than one type of flagellin protein**
- **flagellum are not hollow**
- **hook and basal body difficult to distinguish**
- **more related to Type IV secretions systems**
- **growth occurs at the base, not the end**

# Figure 3.44

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# **Motility**

**Flagellar movement**

**Spirochete motility**

**Twitching motility**

**Gliding motility**

# Motility

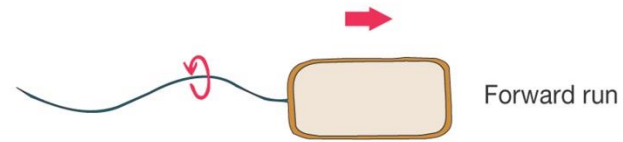
- ***Bacteria* and *Archaea* have directed movement**
- **chemotaxis**
  - **move toward chemical attractants such as nutrients, away from harmful substances**
- **move in response to temperature, light, oxygen, osmotic pressure, and gravity**

# Bacterial Flagellar Movement

- **flagellum rotates like a propeller**
  - **very rapid rotation up to 1100 revolutions/sec**
  - **in general, counterclockwise (CCW) rotation causes forward motion (run)**
  - **in general, clockwise rotation (CW) disrupts run causing cell to stop and tumble**

# Figure 3.45

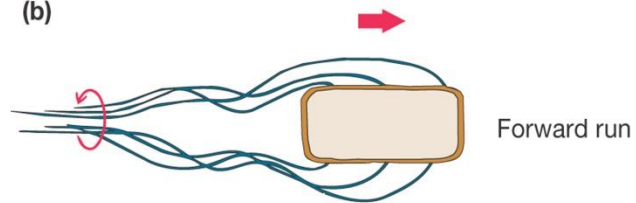
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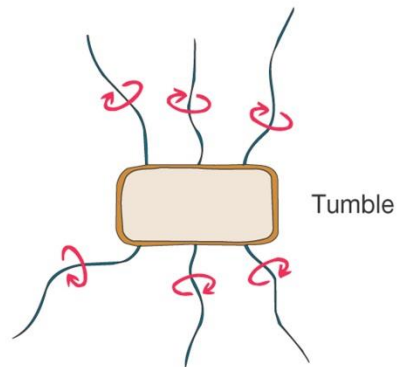
(a)



(b)



(c)



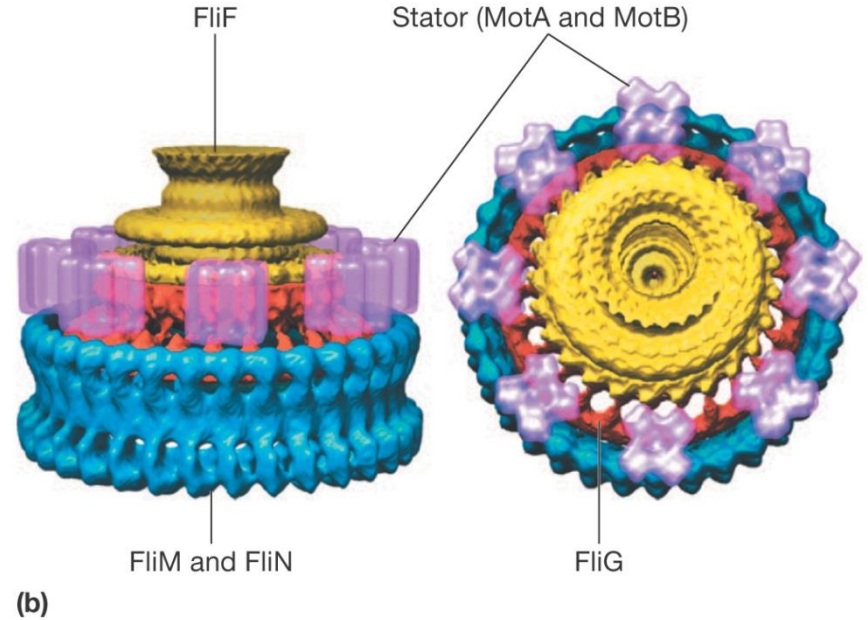
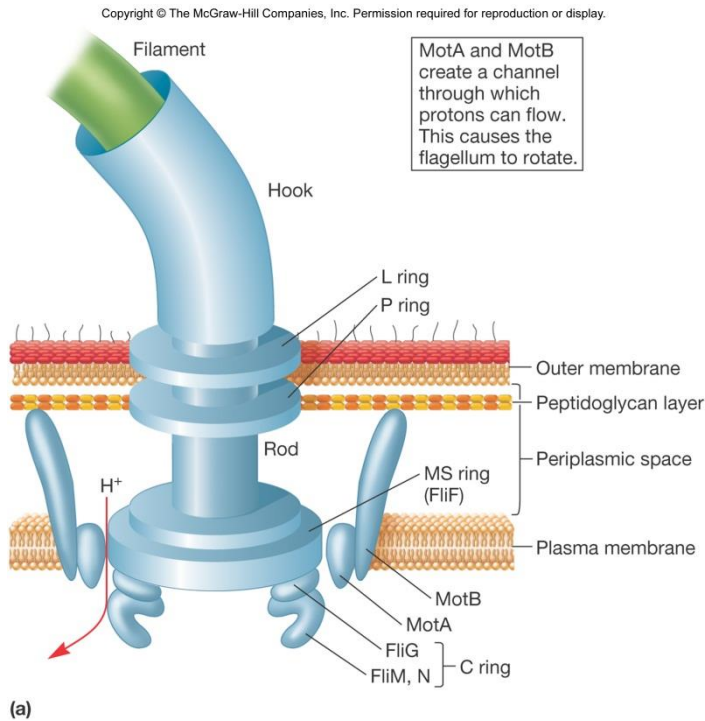
(d)

# Mechanism of Flagellar Movement

- **flagellum is 2 part motor producing torque**
- **rotor**
  - **C (FliG protein) ring and MS ring turn and interact with stator**
- **stator - Mot A and Mot B proteins**
  - **form channel through plasma membrane**
  - **protons move through Mot A and Mot B channels and produce energy through proton motive force**
  - **torque powers rotation of the basal body and filament**

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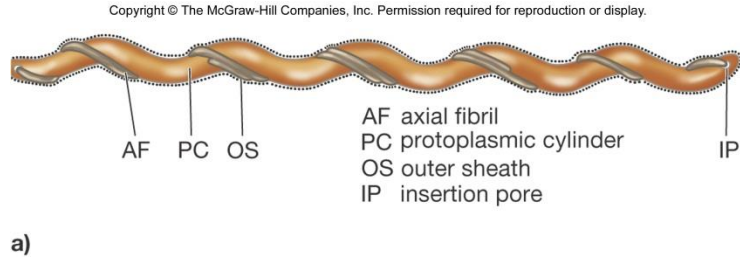
D.R. Thomas, N.R. Francis, C. Xu, and De.J. DeRosier



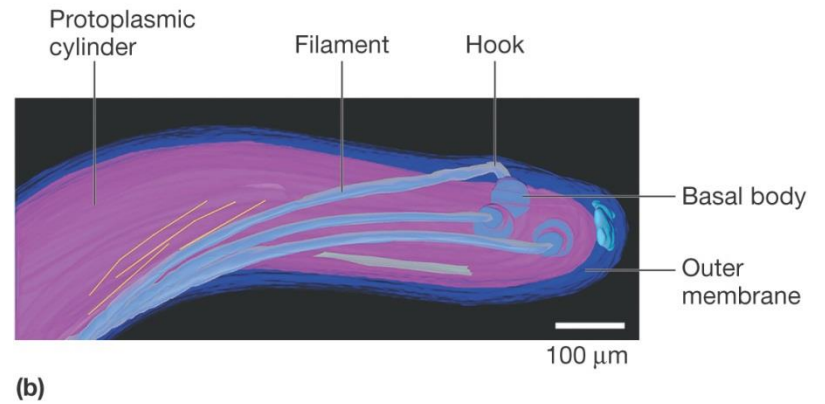
# **Spirochete Motility**

- **multiple flagella form axial fibril which winds around the cell**
- **flagella remain in periplasmic space inside outer sheath**
- **corkscrew shape exhibits flexing and spinning movements**

# Figure 3.47



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Courtesy of Dr. Jacques Izard, Forsyth Institute, Boston

# **Twitching and Gliding Motility**

- **may involve Type IV pili and slime**
- **twitching**
  - **pili at ends of cell**
  - **short, intermittent, jerky motions**
  - **cells are in contact with each other and surface**
- **gliding**
  - **smooth movements**

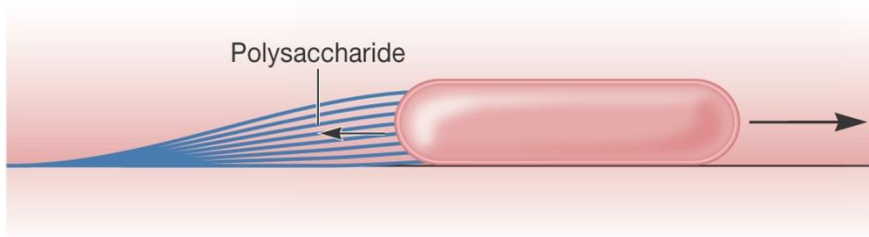
# ***Myxococcus xanthus***

## **Movement**

- **social**
  - **Type IV pili move together in large groups of cells**
- **adventurous (Gliding)**
  - **alime released moves cell forward**
  - **adhesion complexes move in track provided by cytoskeleton**

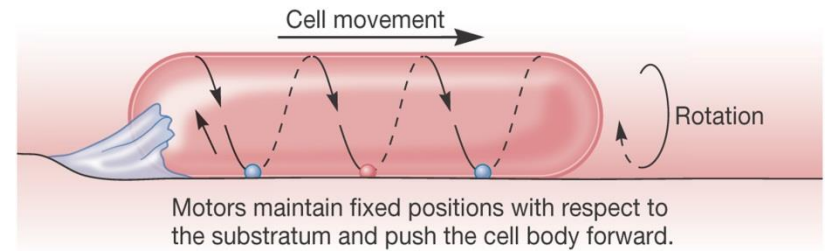
# Figure 3.48

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(a)

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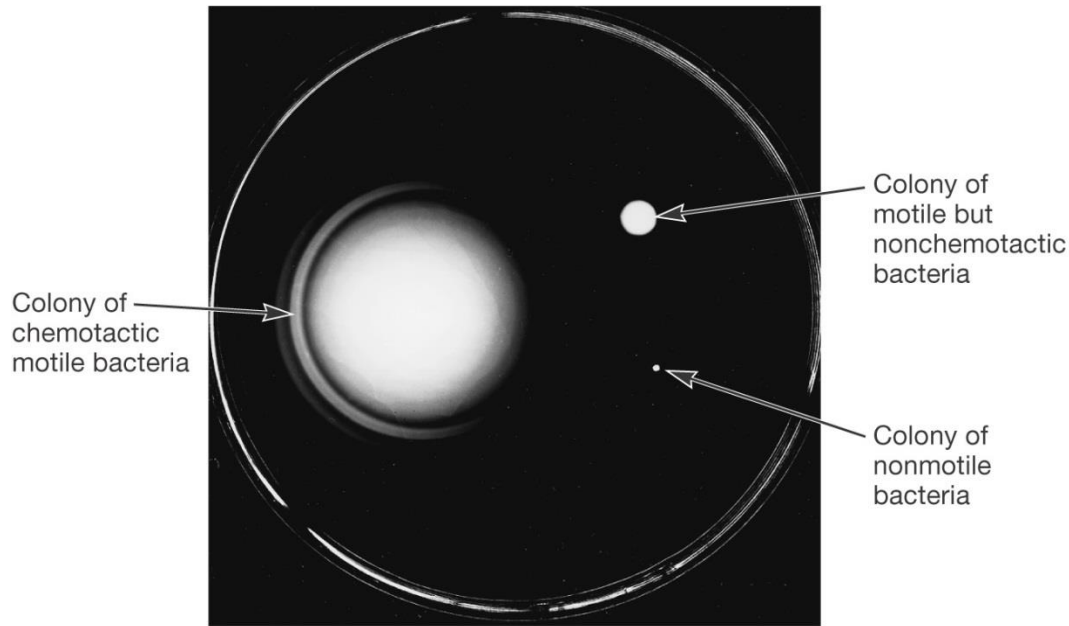
(b)

# Chemotaxis

- **movement toward a chemical attractant or away from a chemical repellent**
- **changing concentrations of chemical attractants and chemical repellents bind chemoreceptors of chemosensing system**

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(a) Positive chemotaxis



(b) Negative chemotaxis

Courtesy of Dr. Julius Adler

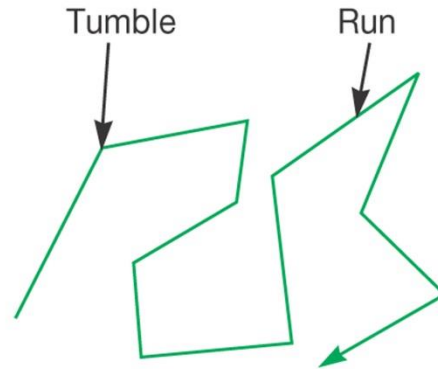
# Chemotaxis

- **in presence of attractant (b) tumbling frequency is intermittently reduced and runs in direction of attractant are longer**
- **behavior of bacterium is altered by temporal concentration of chemical**
- **chemotaxis away from repellent involves similar but opposite responses**



# Figure 3.50

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(a)



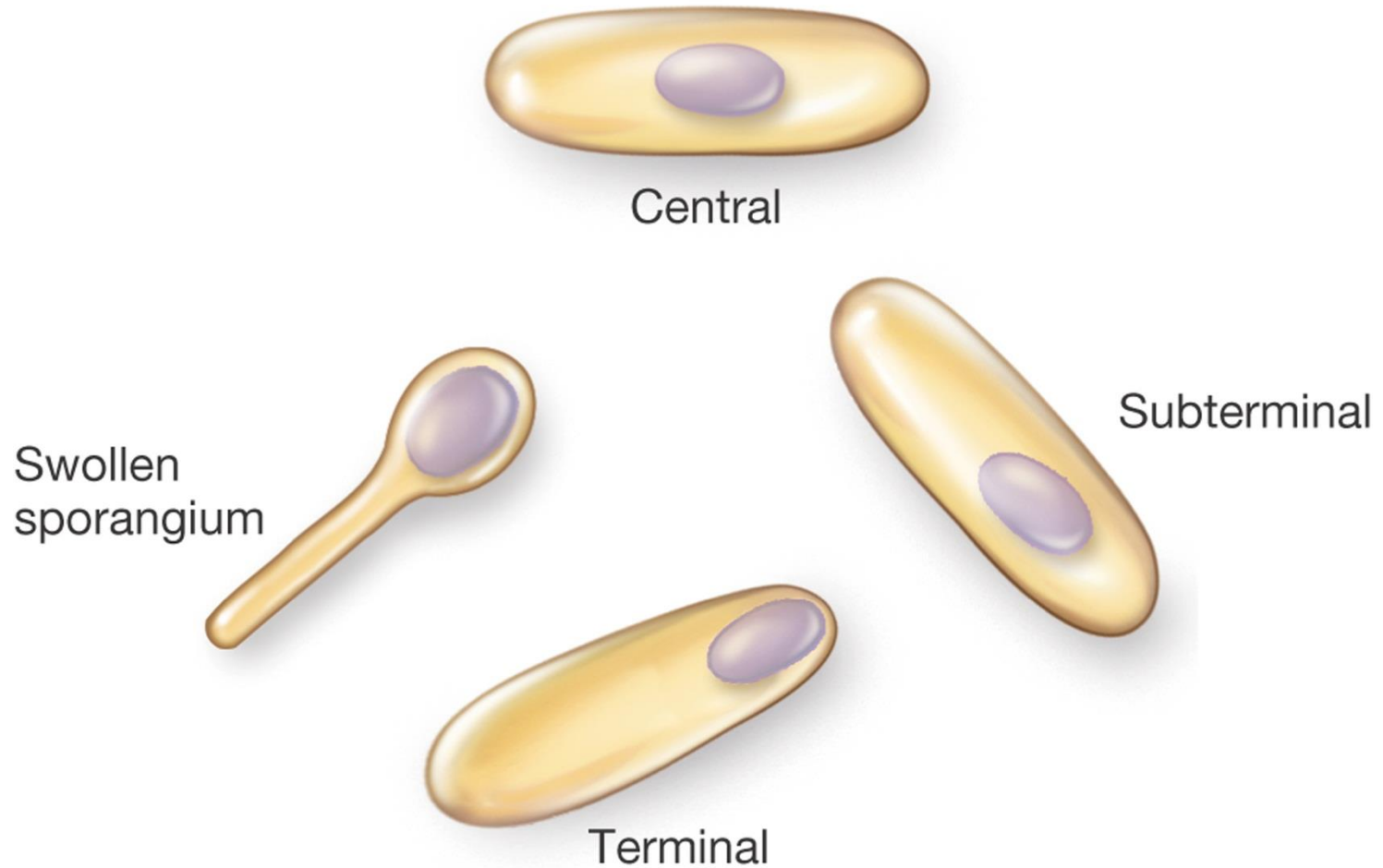
(b)

# **The Bacterial Endospore**

- **complex, dormant structure formed by some bacteria**
- **various locations within the cell**
- **resistant to numerous environmental conditions**
  - **heat**
  - **radiation**
  - **chemicals**
  - **desiccation**

# Figure 3.51

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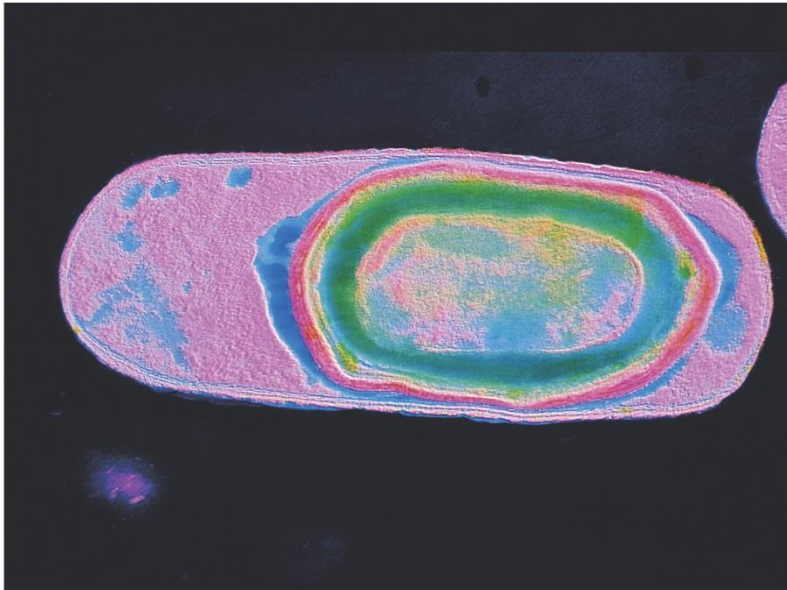


# Endospore Structure

- **spore surrounded by thin covering called exosporium**
- **thick layers of protein form the spore coat**
- **cortex, beneath the coat, thick peptidoglycan**
- **core has nucleoid and ribosomes**

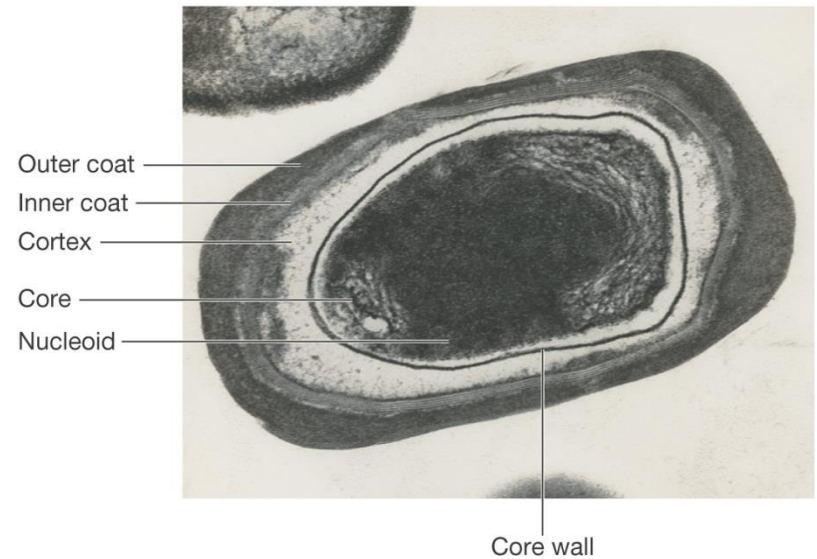
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# **What Makes an Endospore so Resistant?**

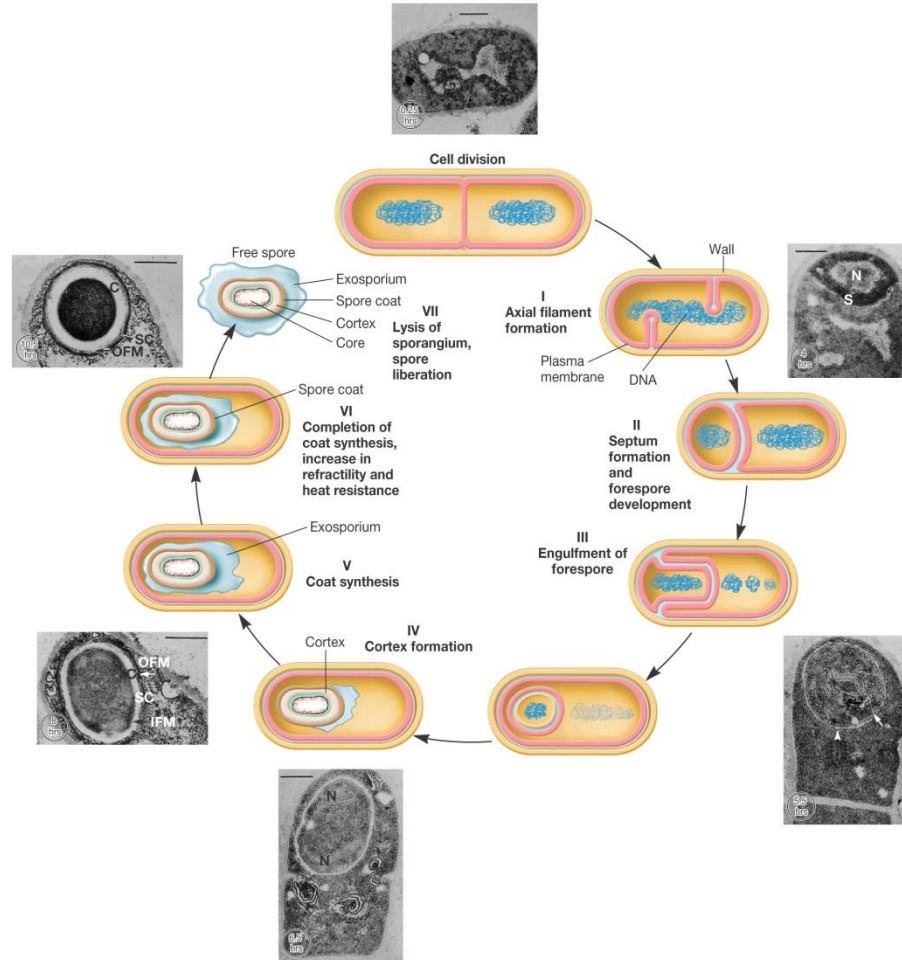
- **calcium (complexed with dipicolinic acid)**
- **small, acid-soluble, DNA-binding proteins (SASPs)**
- **dehydrated core**
- **spore coat and exosporium protect**

# Sporulation

- **process of endospore formation**
- **occurs in a hours (up to 10 hours)**
- **normally commences when growth ceases because of lack of nutrients**
- **complex multistage process**

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From A.N. Barker et al. (Eds.), Spore Research, 1974, pages 161-174, 1971 Academic Press

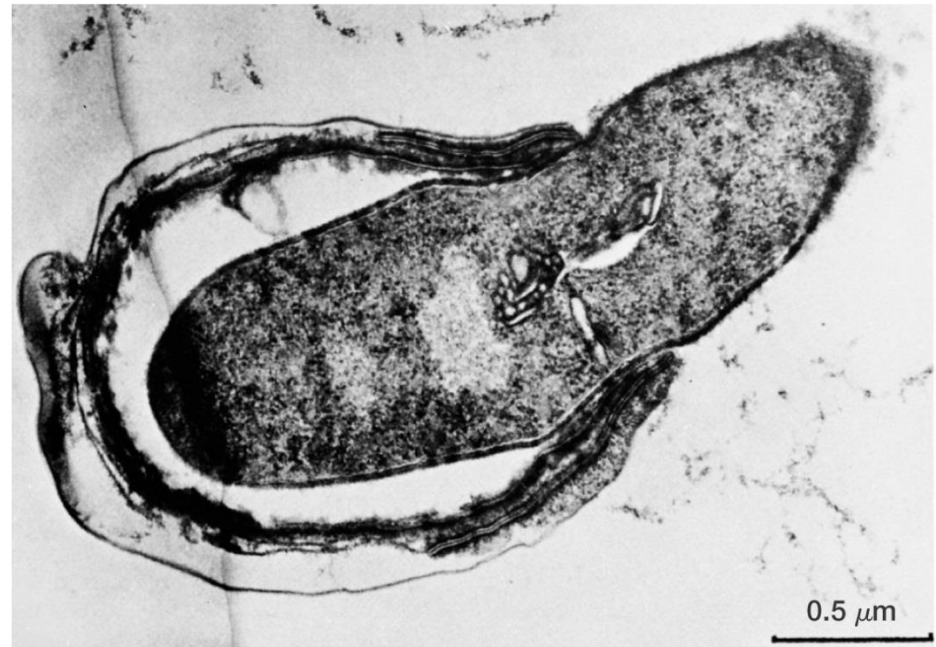


# Germination

- **transformation of endospore into vegetative cell complex, multistage process**

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# Formation of Vegetative Cell

- **activation**
  - prepares spores for germination
  - often results from treatments like heating
- **germination**
  - environmental nutrients are detected
  - spore swelling and rupture of absorption of spore coat
  - loss of resistance
  - increased metabolic activity
- **outgrowth - emergence of vegetative cell**